

# Insights into Phosphatidylethanol (PEth) 16:0/18:1 Formation and Degradation Analysis by Supercritical Fluid Chromatography-Tandem Mass Spectrometry

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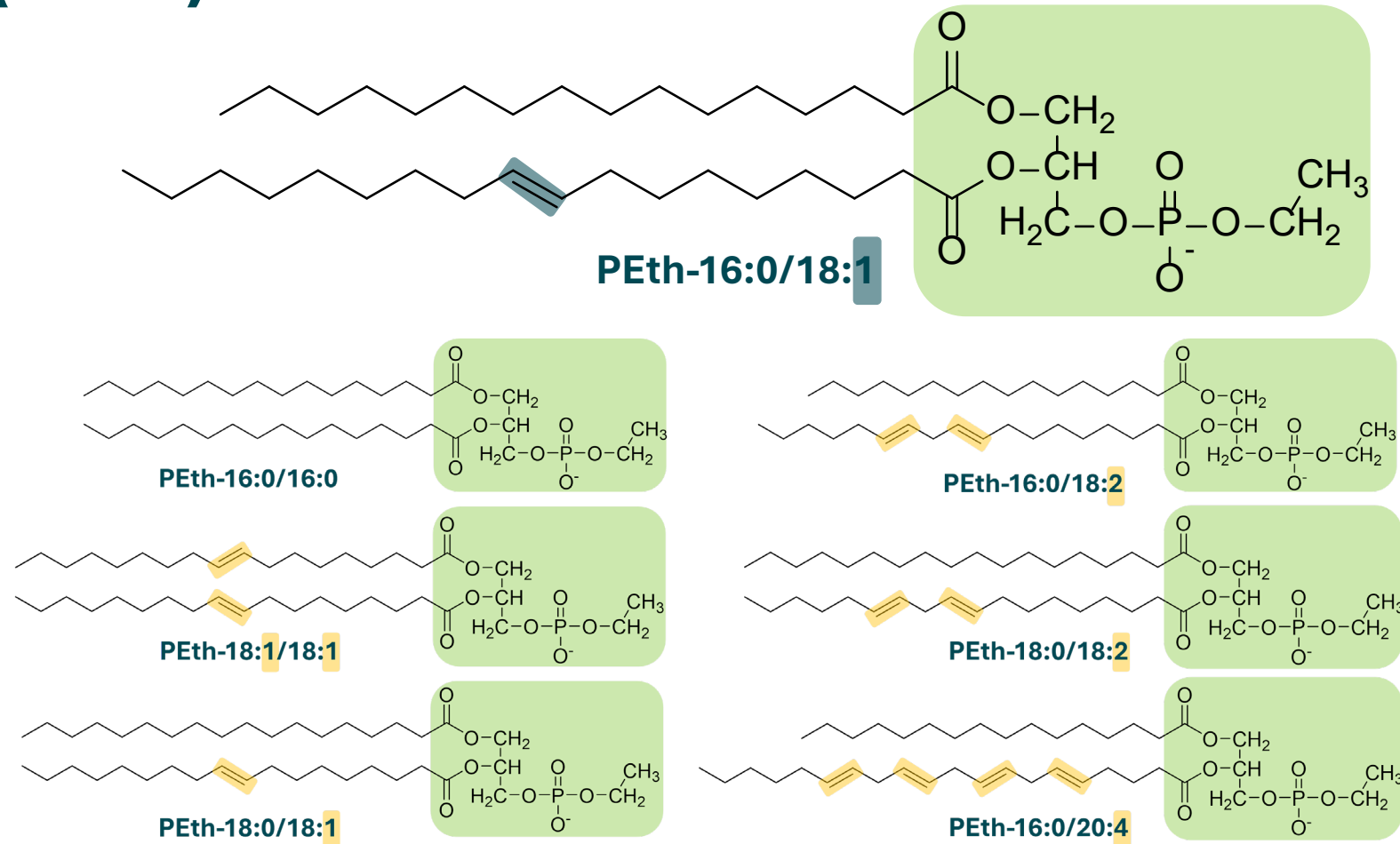
# Disclaimer

- The views and opinions expressed in this presentation are solely mine, and do not represent any official views or opinions of the American Academy of Forensic Sciences.
- I have no financial or conflicts of interest to disclose.



# Phosphatidylethanol (PEth)

- Abnormal phospholipids formed in erythrocyte membranes in the presence of ethanol
- PEth 16:0/18:1 predominant homologue
- Highly correlated to alcohol intake, even at low doses
- Relatively insensitive to unintentional alcohol exposure
- Long detection window (~12 days) after single drinking event





# PEth Testing

- Detect changes in drinking behavior earlier and more accurately compared to other biomarkers (e.g. EtG, EtS etc)
  - Useful for abstinence monitoring, diagnosing alcohol use disorder
- Postmortem (PM) PEth provides information on alcohol consumption in the weeks preceding death
  - Useful in determining chronic drinking in cases where alcohol is absent
- 2022 Consensus of Basel
  - *Harmonize PEth analysis and interpretation on a global scale*

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LETTER TO THE EDITOR

WILEY

**Consensus for the use of the alcohol biomarker phosphatidylethanol (PEth) for the assessment of abstinence and alcohol consumption in clinical and forensic practice (2022 Consensus of Basel)**

**PEth 16:0/18:1**

**concentration cutoff**

**Interpretation**

<20 ng/ml

Compatible with abstinence or low alcohol consumption

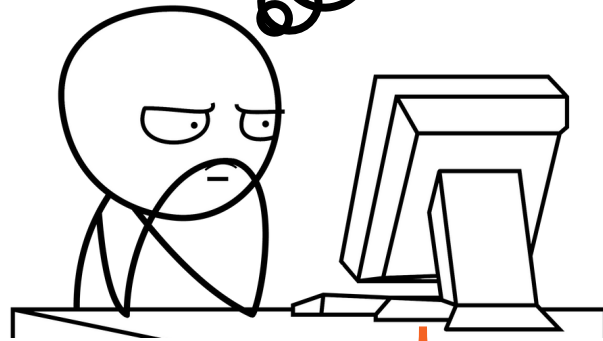
≥20 ng/ml but <200 ng/ml

Alcohol consumption

≥200 ng/ml

Strongly suggestive of chronic excessive alcohol consumption

- 
- **How is PEth formation and degradation impacted during algor mortis?**



**Drinking event**

**+**

**Time of death**

**Blood collection and storage at -80°C**

- Risk of post-sampling formation of PEth in blood containing ethanol
  - Dried blood spots
  - PLD inhibitors (FIPI,  $\text{NaVO}_3$ ,  $\text{Na}_2\text{WO}_4$ )



# Research Objectives

1. Develop and validate the first SFC-MS/MS method to quantify PEth 16:0/18:1 in PM blood
  - *per ANSI/ASB 036 recommendations and in-house validation protocols*



# Liquid-liquid extraction

- Weigh *blood (0.25 g)*
- Add *250  $\mu$ L of pH 9 borate buffer (0.6M)*
- Add *2 mL of 80:20 (v/v) heptane:2-propanol*
- Centrifuge (4000 rpm, 10 minutes)
- Freeze at  $-80^{\circ}\text{C}$  for 15 min
- Decant organic layer
- Evaporate to dryness at  $40^{\circ}\text{C}$  under nitrogen (1.5 mL/min, 15 min)
- Reconstitute in *200  $\mu$ L methanol*



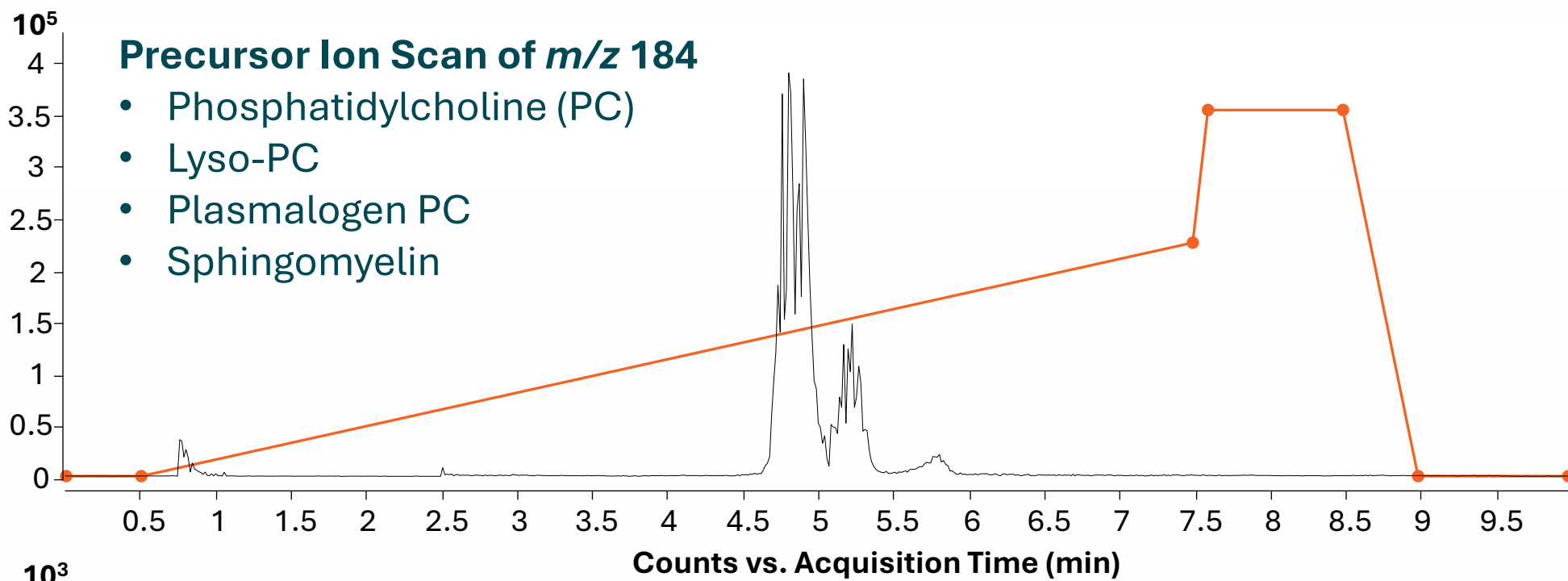
# Method Parameters

Agilent 1260 Infinity II SFC control module and 1260 Infinity II Liquid Chromatograph	
<b>Column and Temperature (°C)</b>	Waters Corp Torus DEA column (1.7 $\mu$ m, 3 mm X 100 mm) at 50°C
<b>Mobile Phase</b>	<b>A:</b> Supercritical CO <sub>2</sub> <b>B:</b> 0.1% ammonium acetate in 95:5 methanol: water
<b>Flow Rate</b>	1.0 mL/min
<b>Injection volume</b>	2 $\mu$ L

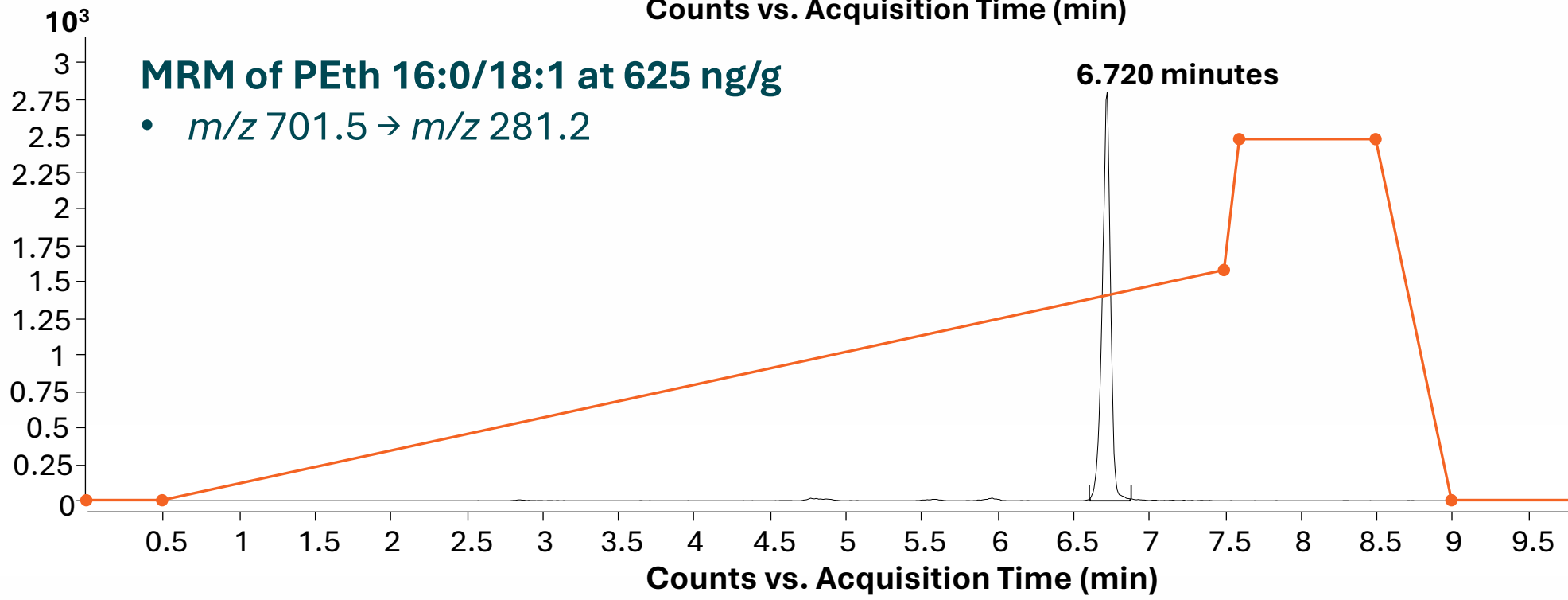
Agilent Ultivo Triple Quadrupole Mass Spectrometer	
<b>Ionization type</b>	Electrospray Ionization (ESI) in negative mode
<b>Acquisition Type</b>	Multiple Reaction Monitoring
<b>PEth 16:0/18:1</b>	<b>Quantifier:</b> $m/z$ 701.5 $\rightarrow$ $m/z$ 281.2; <b>Qualifier:</b> $m/z$ 701.5 $\rightarrow$ $m/z$ 255.2
<b>PEth 16:0/18:1-d<sub>5</sub></b>	<b>Quantifier:</b> $m/z$ 706.5 $\rightarrow$ $m/z$ 281.2







Gradient	
Minutes	% B
0	1
0.5	1
7.5	45
7.6	70
8.5	70
9	1
10	1



Parameters	Validation Procedure	Results
	*LQC: 20 ng/g, MQC: 200 ng/g, HQC: 2000 ng/g	
Recovery (%)	Evaluated at LQC and HQC concentrations in 10 sources	59% (LQC), 48% (HQC)
Linearity	Least square regression and residual plot with 6 non-zero calibrators over 8 days	Linear 1/x between 10 – 2500 ng/g
LOD/LLOQ	Lowest calibrator extracted in triplicate with 3 autopsy sources over 3 runs (n=9)	LOD administratively set at LLOQ (10 ng/g)
Bias	Extracted in triplicate over 8 days	-16.4 to 17.7% (LQC), -8.8 to 12.5 % (MQC), -15.2 to 10.4% (HQC)
Within-run %CV		2.9 to 17.1% (LQC), 0.7 to 7.6% (MQC), 1.3 to 10.5% (HQC)
Between-run %CV	Combined replicate pools of triplicate extracted QCs over 8 days	8.3% (LQC), 5.6% (MQC), 5.7% (HQC)
Matrix Effects	Post-extraction addition with 10 sources (n=20)	20% (LQC), 1% (HQC)
Interference studies	Monitor (1) Negatives, (2) neat homologues (PEth 18:0/18:1, 16:0/16:0, 18:1/18:1, 18:0/18:2, 16:0/18:2, and 16:0/20:4), (3) 10 'blank' autopsy sources, (4) ULOQ with no internal standard	No interferences
Dilution integrity	1:5 dilution of HQC (n=5)	-17.6 % (Bias), 2.3% (Precision)
Carryover	Re-injecting extracted blank after highest calibrator (2500 ng/g) over 8 runs	No carryover
Processed sample stability	Peak area of PEth/PEth-d <sub>5</sub> after 72 hrs at 8°C for 20 ng/g (LQC) and 2000 ng/g (HQC) concentration	≤ 15% signal decrease from the t <sub>0</sub> response



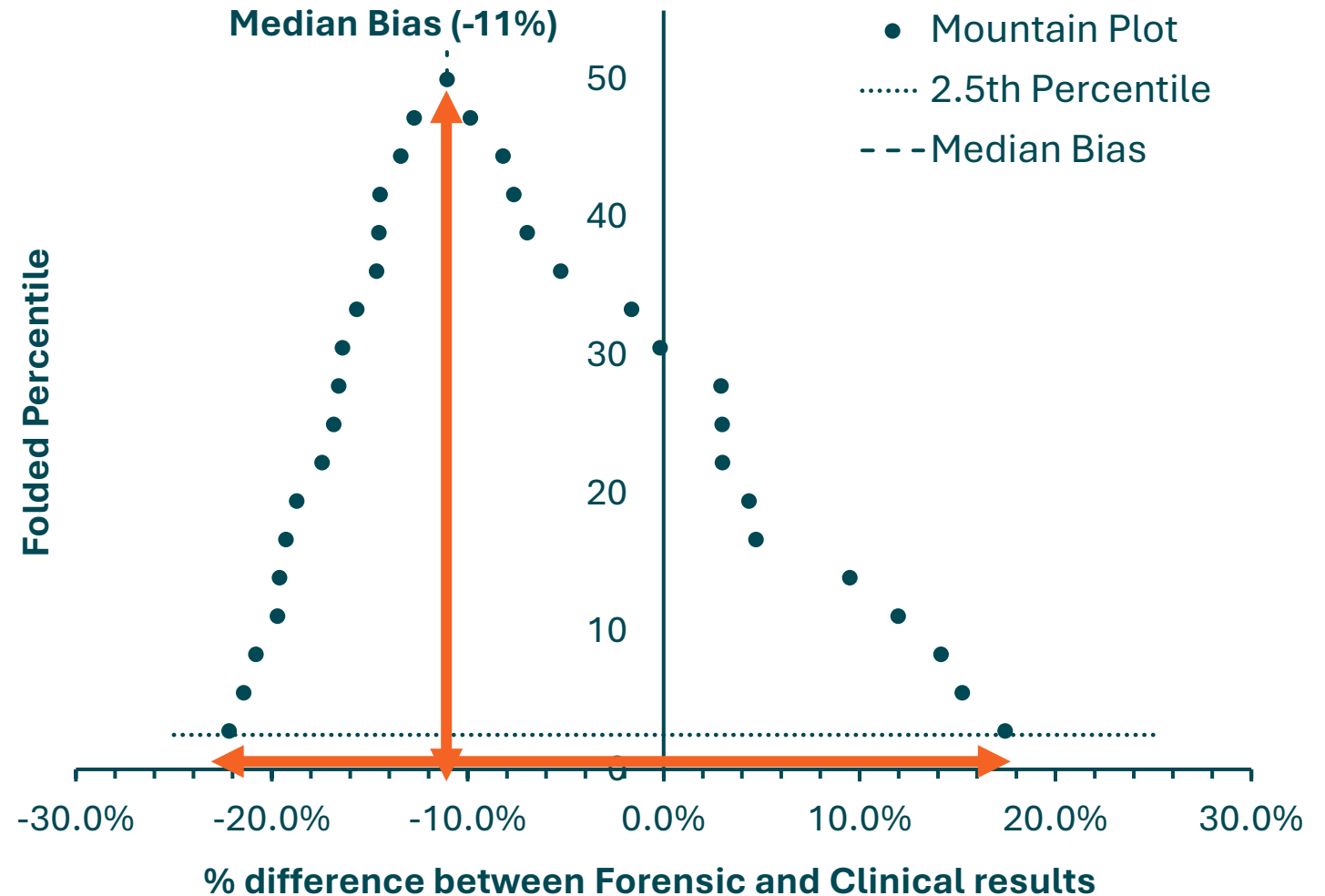
## Proof-of-concept:

- 35 autopsy samples ranging between 32.6 to 2476 ng/g
- Mountain plot to illustrate differences between clinical and forensic method

**Clinical:** *SPE and LC-MS/MS*

**Forensic:** *LLE and SFC-MS/MS*

- Forensic SFC-MS/MS method generally quantified lower PEth levels than clinical LC-MS/MS method
  - Changes in PM blood could contribute to differences



# Research Objectives

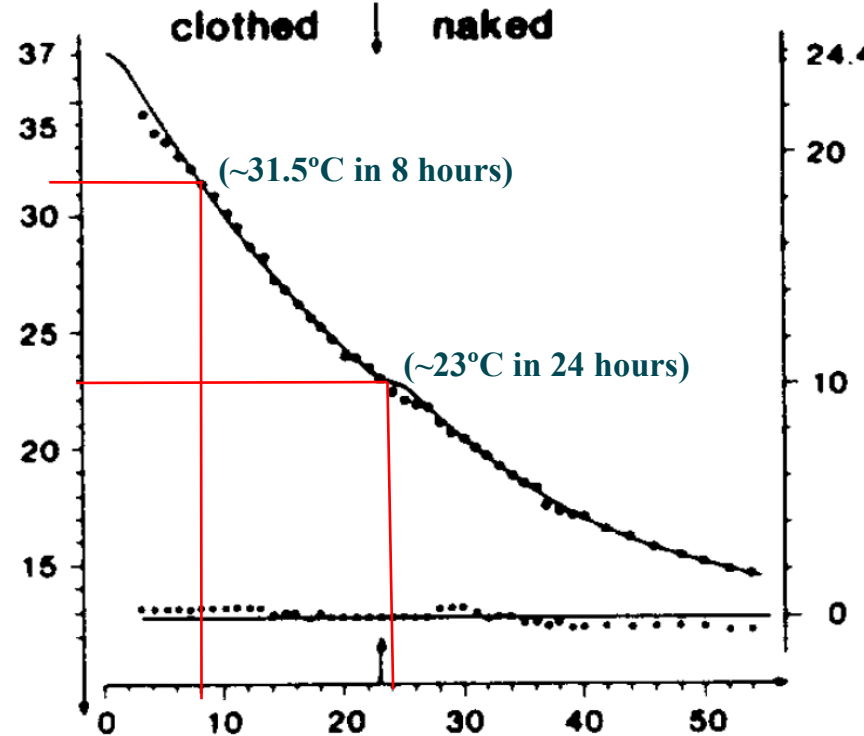


1. Develop and validate the first SFC-MS/MS method to quantify PEth 16:0/18:1 in PM blood
  - *per ANSI/ASB 036 recommendations and in-house validation protocols*
2. Develop a simple *in vitro* model to simulate PEth 16:0/18:1 formation and degradation in the first 48 hours postmortem

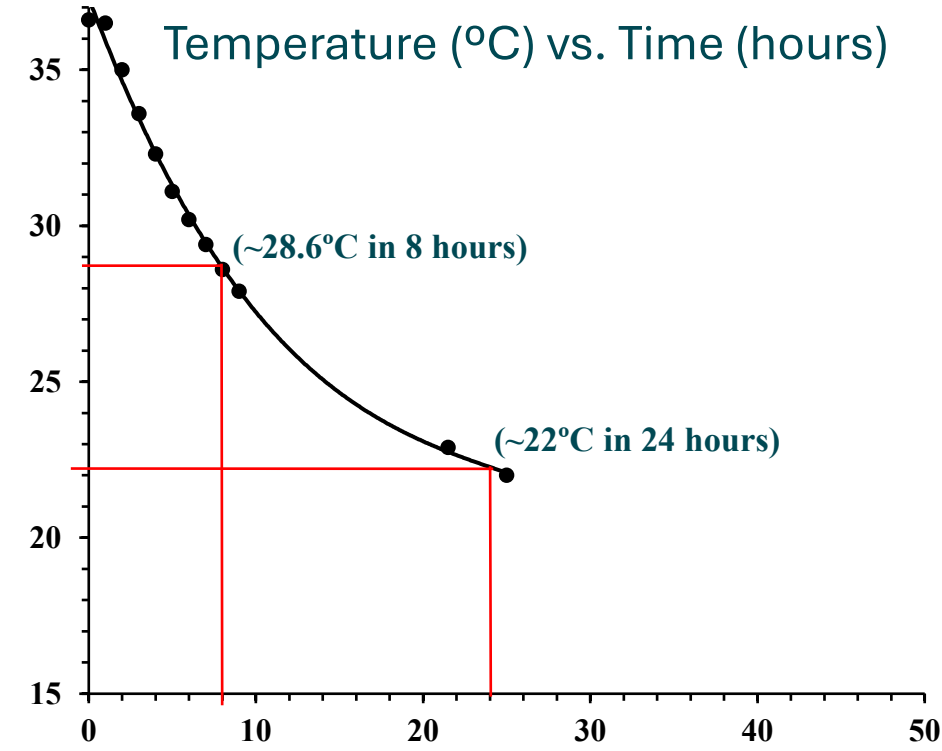
# Henssge's Nonogram

- *Cooling of body as sigmoidal function*
- *Initial plateau before cooling according to Newton's law of cooling*
- *Simplified model offers fair estimates*

**Henssge's cooling curve**



**in vitro model**



**Assumptions:**

1. *Core body temp. at time of death is approx.  $37^{\circ}\text{C}$*
2. *Body temperature PM would reach ambient ( $22^{\circ}\text{C}$ ) in 24 hours*



BAC 0 g/L and heparinized

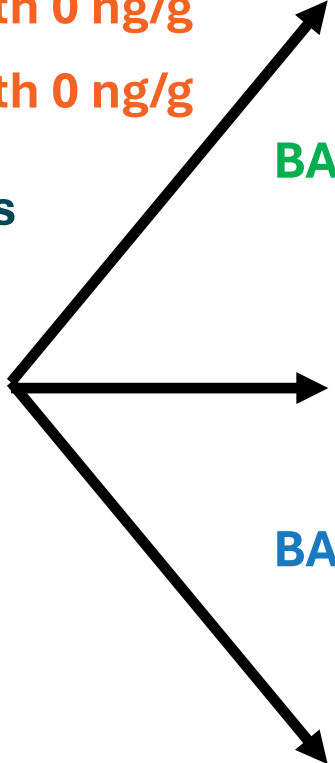
Source 1: PEth 14.4 ng/g

Source 2: PEth 31.0 ng/g

Source 3: PEth 0 ng/g

Source 4: PEth 0 ng/g

4 sources



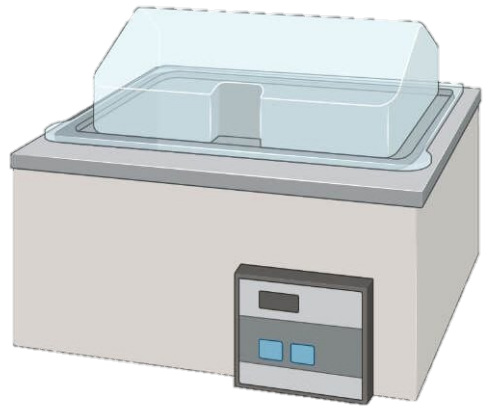
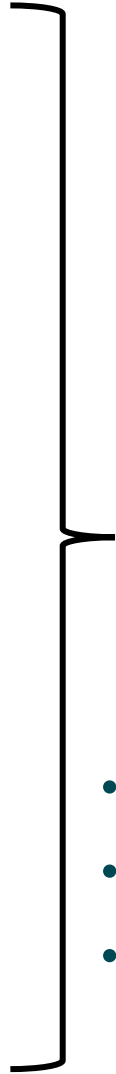
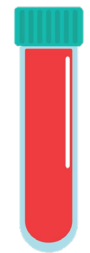
BAC 1.5 g/L



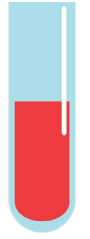
BAC 3.0 g/L



BAC 0.0 g/L



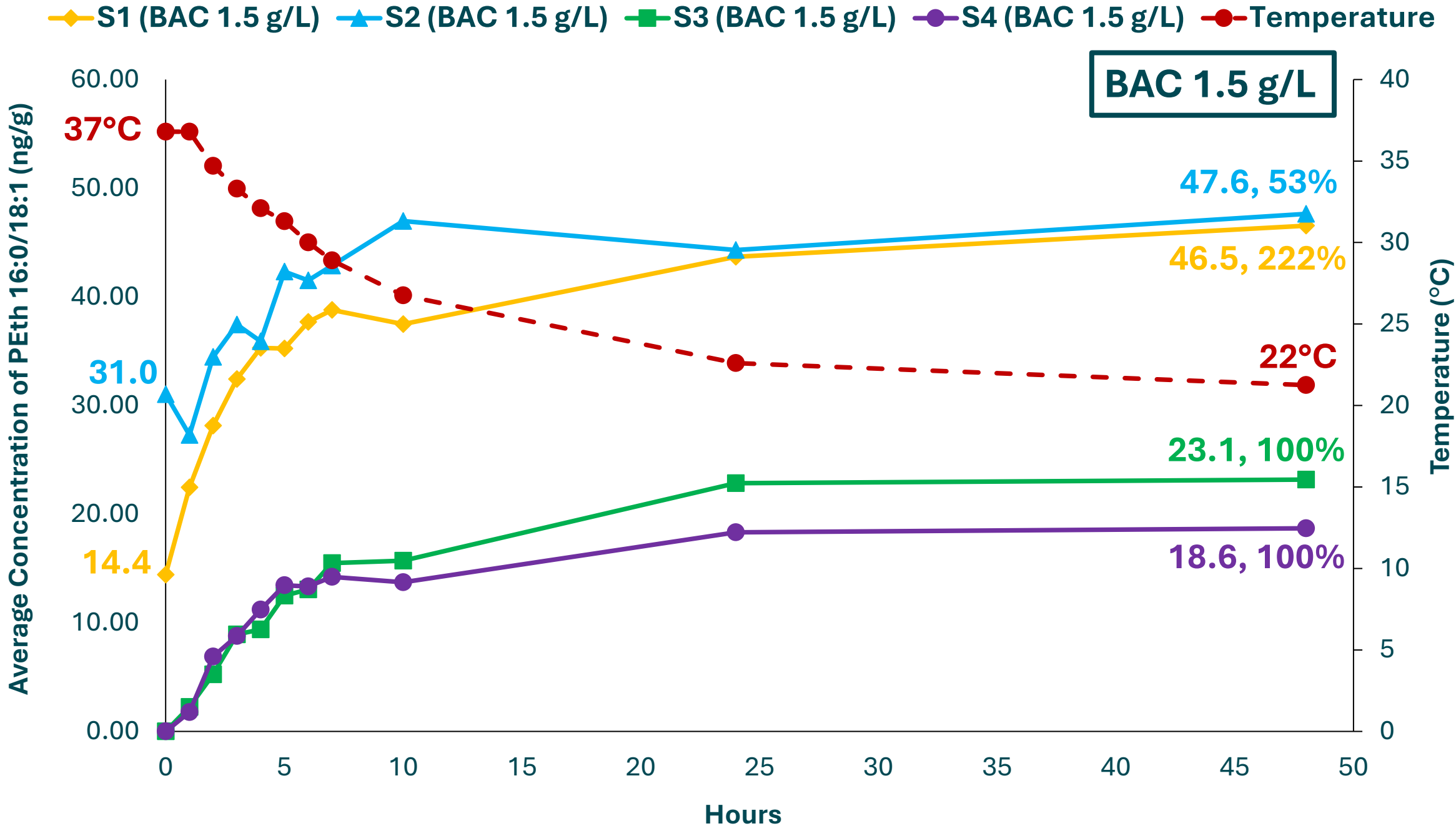
- Set at 37°C for 1 hour
- Turned off with lid on
- Left to cool back down to room temp. (22°C) over time



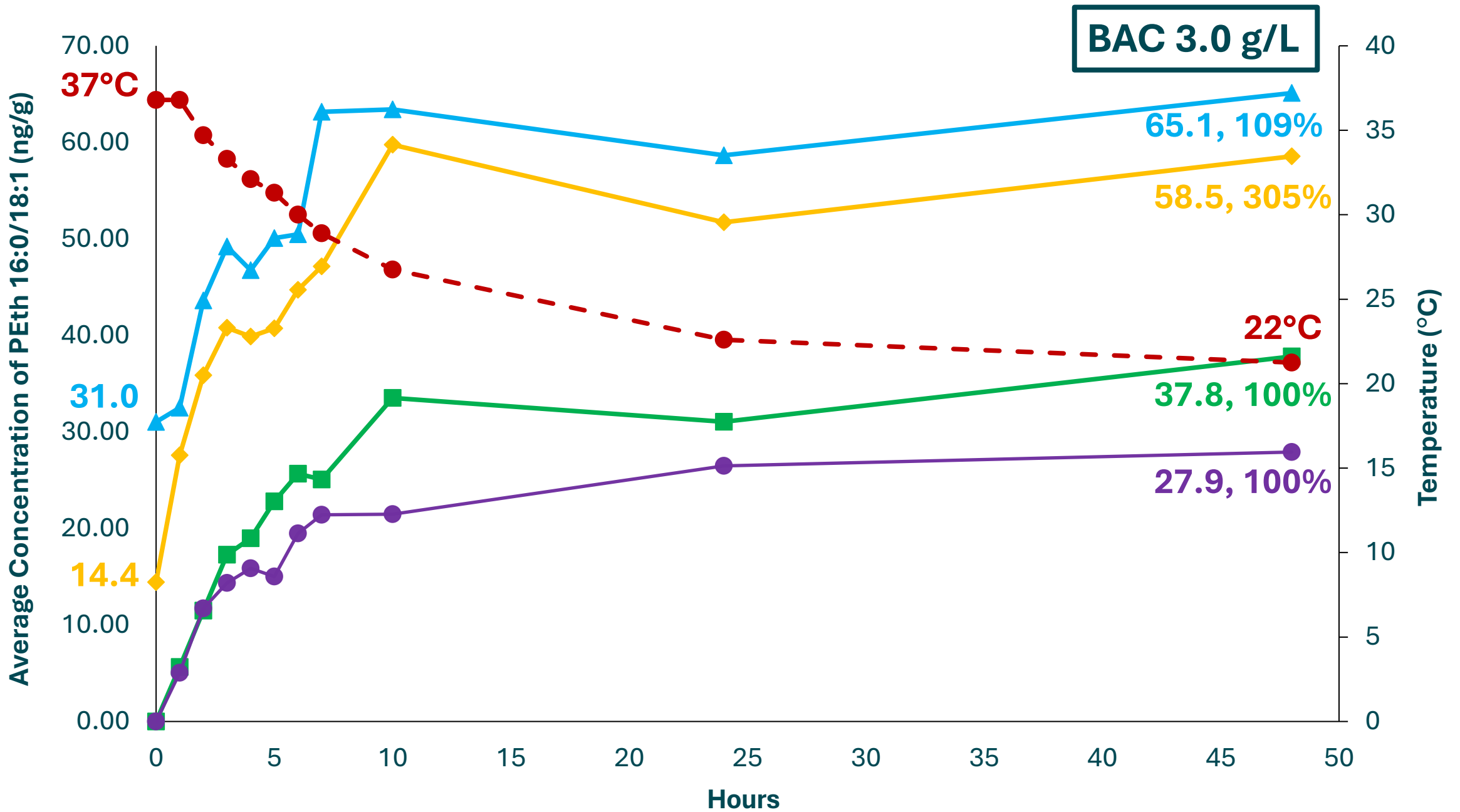
x2

*Aliquot in duplicate per each source*

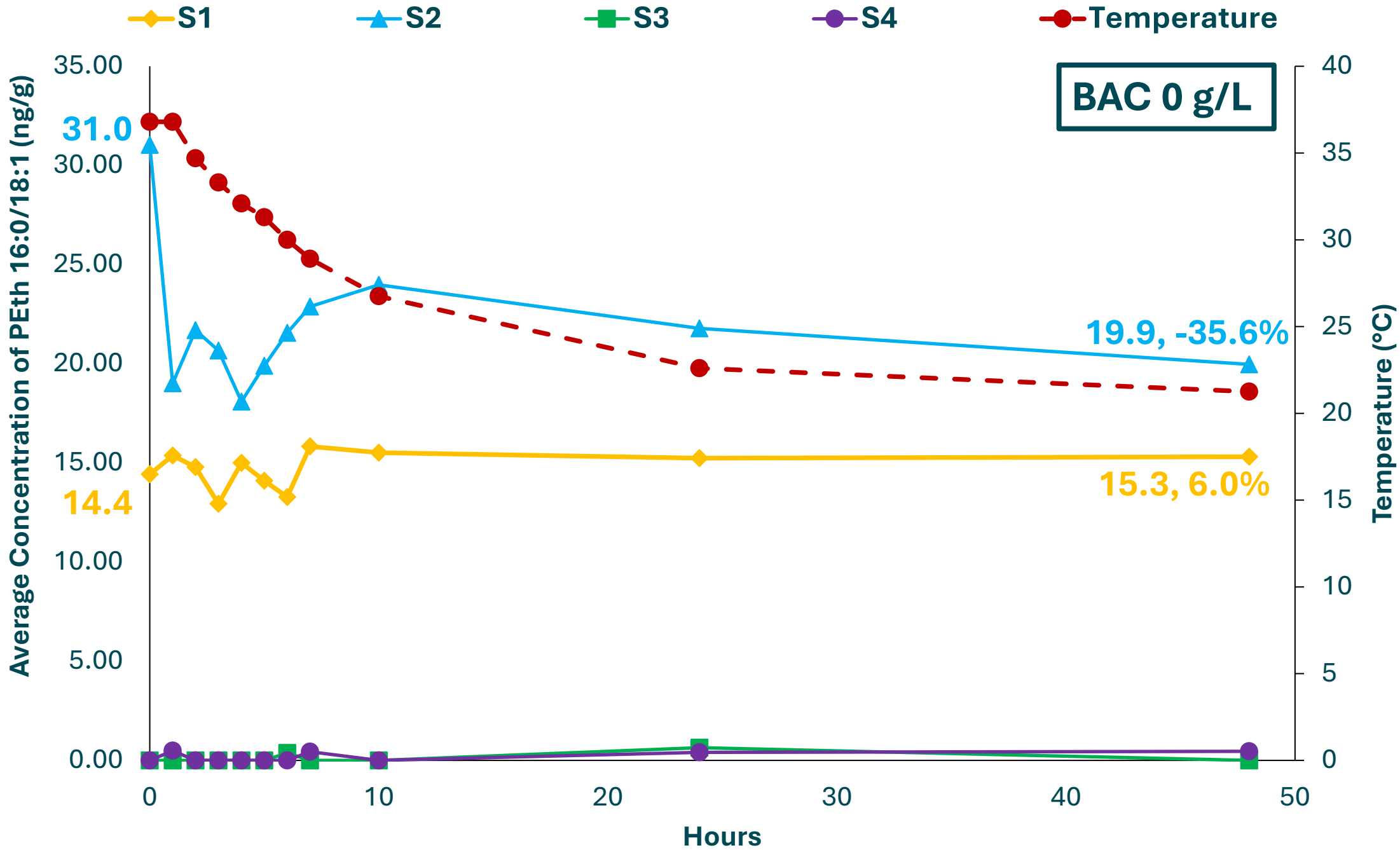
- Once an hour for 7 hours
- After 24 hours
- After 48 hours
- Recorded temperature during each sampling



—◆— S1 (BAC 3.0 g/L) —▲— S2 (BAC 3.0 g/L) —■— S3 (BAC 3.0 g/L) —●— S4 (BAC 3.0 g/L) —●— Temperature









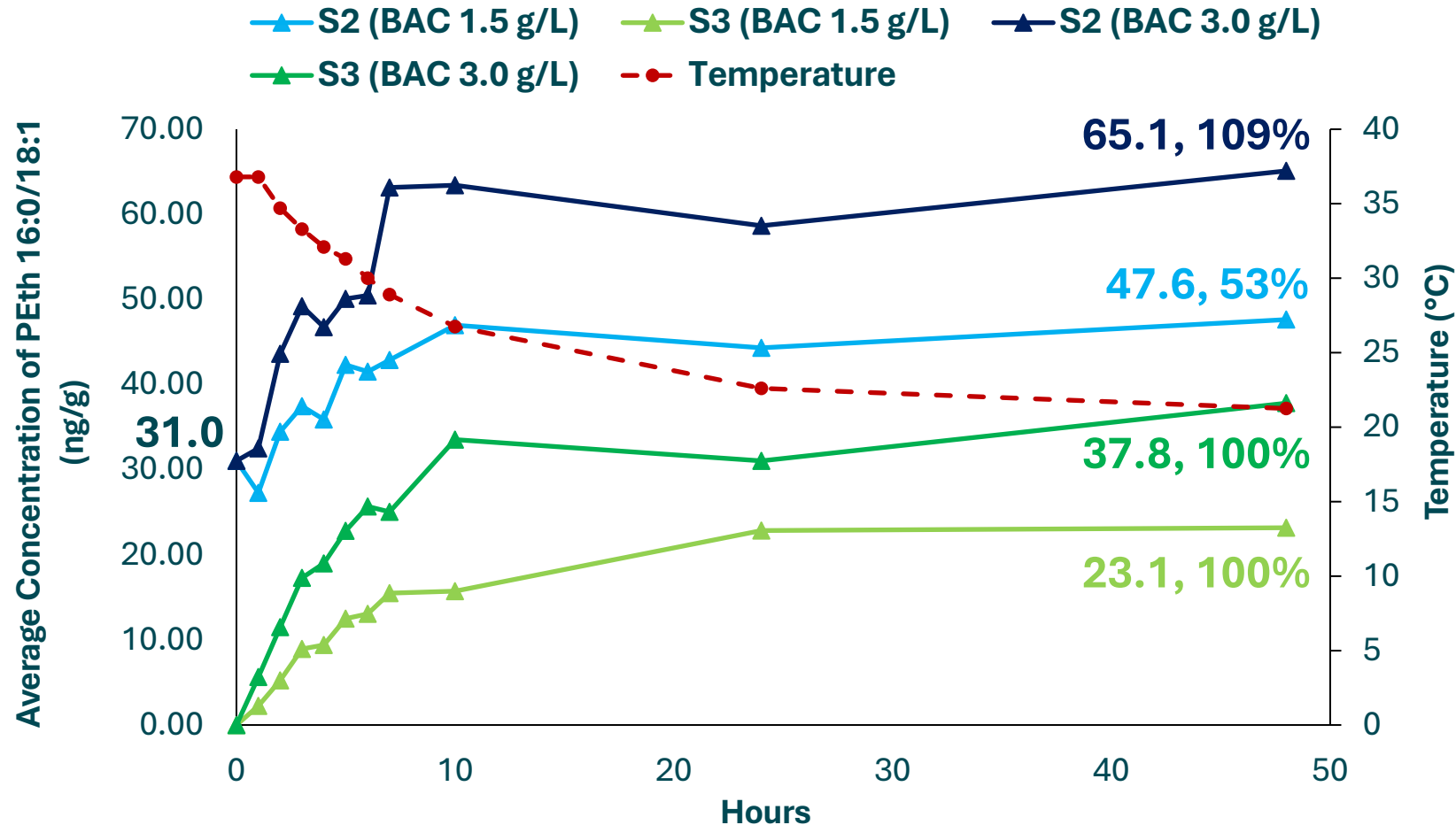
# Results

1. Formation occurs and decreases at lower temperatures, levels before reaching room temperature, and with minimal PEth formation after 10 hours
  - Simultaneous formation and degradation of PEth 16:0/18:1
2. PEth formation occurs in an ethanol concentration-dependent manner
  - Statistically significant ( $\alpha=0.05$ ) difference between BAC 1.5 g/L and 3.0 g/L groups
3. Results from our study aligns with previous findings from Schröck *et al.* (2018) study of *in vitro* PEth formation at 37°C for 7 hours



# Conclusions

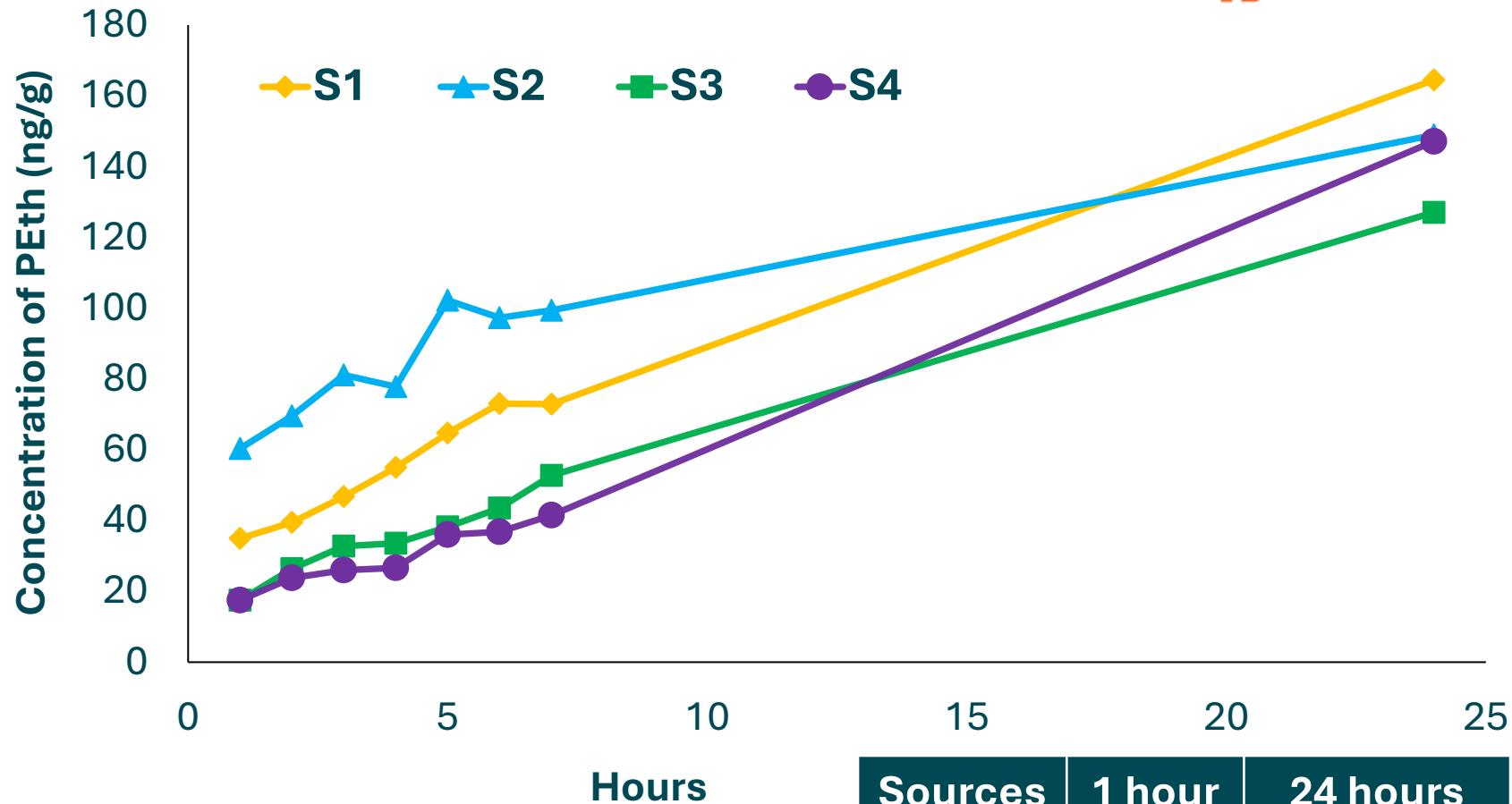
- At temperatures below 37°C, phospholipase D remains active at a lower rate and continues to produce PEth
  - Interindividual variability in PEth formation
- Temperature is a rate-limiting factor
  - Limited BAC decline (5.2-22.6% decrease)





# Follow-Up

- Same 4 blood sources fortified to BAC 3.0 g/L and incubated at 37°C for 24 hours
- Continuous PEth formation with higher final PEth concentrations
- Consistent with findings by Schröck et al. (2018)



Sources	1 hour (ng/g)	24 hours (ng/g)
1	34.9	164.4
2	60.3	148.9
3	17.4	127.0
4	17.4	147.1



# Summary

- Measured PEth not an accurate representation of exact levels at time of death
- Cases with high BAC and delayed sample collection are more prone to PM PEth formation
  - Extra considerations needed if decedent's body has been left at elevated temperatures
- *in vitro* modelling cannot be directly extrapolated to fully replicate complex physiological processes that occur within the human body
  - Larger sample size needed
- Findings reinforce key considerations in the 2022 Consensus of Basel on PEth interpretation
  - Necessity to account for relevant, case-specific factors

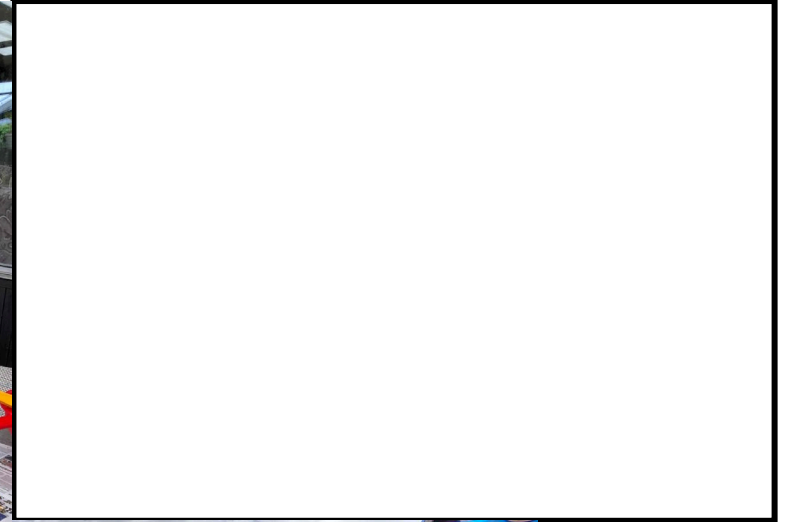


# Acknowledgements

- Markus Roman
- Dr. Britni Skillman
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- Dr. Robert Kronstrand



RÄTTSMEDICINALVERKET



## Coming soon on the Journal of Analytical Toxicology!

**Title:** Quantification of Phosphatidylethanol 16:0/18:1 in blood using Supercritical Fluid Chromatography-Tandem Mass Spectrometry





# Questions?

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