

Lucilia mexicana and its Population Structure in Texas, USA

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Forensic Entomology

Application of insects to civil and criminal investigations

Urban

PEST CONTROL SERVICE



Stored Products

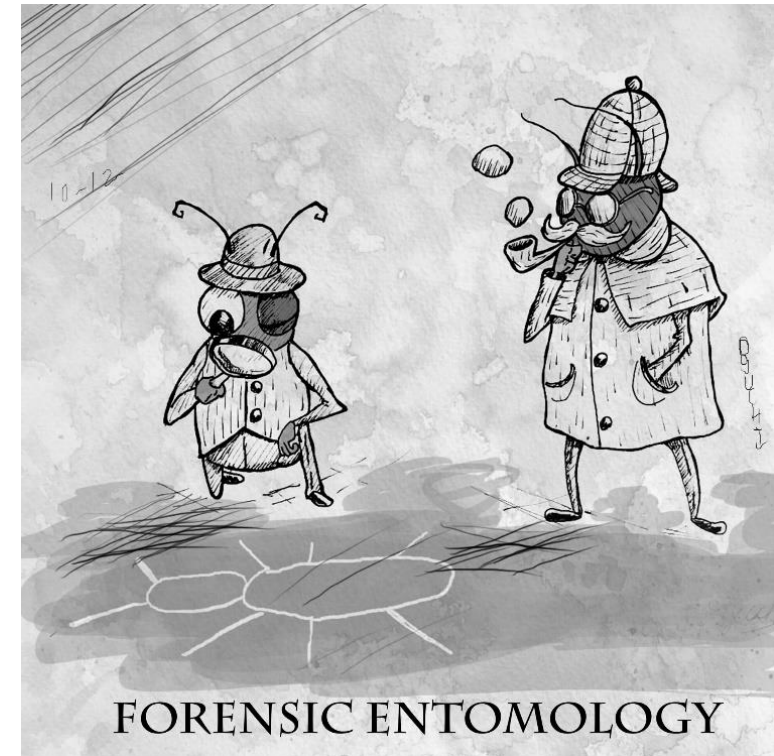


Medico-legal



Insects in Medico-Legal Investigations

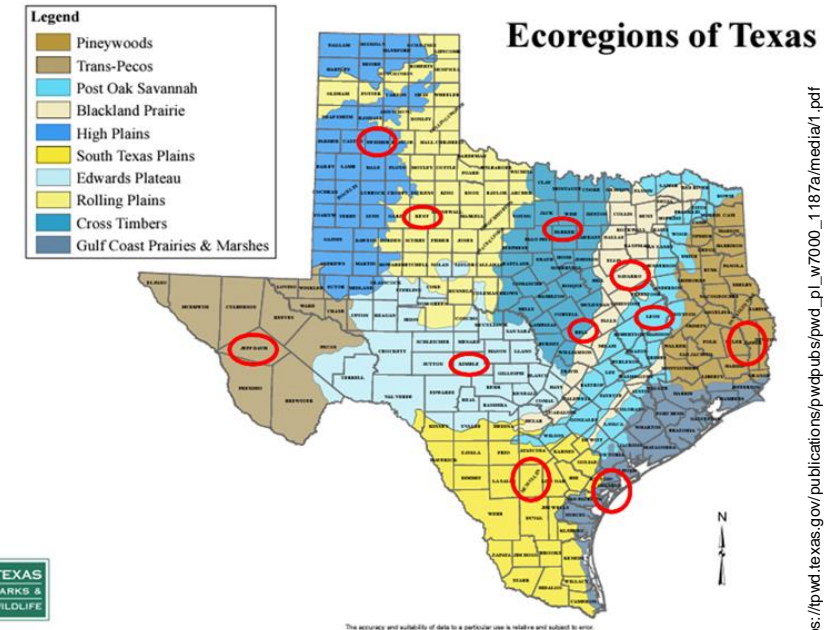
- Insects are the first to arrive at a body
 - Most common flies and beetles
- Insects are an important tool¹
 - Post-mortem Interval (PMI) Estimation
 - **DNA Analysis**
- Entomological evidence
 - Geographical region
 - Season



Forensic Entomology (Student Magazine Illustration), Oguchi Anyaë. (n.d.).
<https://ogu-chiart.artstation.com/projects/3oLm22> (accessed February 5, 2021).

Lucilia mexicana

- Southwestern United States distribution^{2,3}
- First colonizer of mammalian carrion in Texas⁴
 - Myiasis-infestation of living tissue by maggots
- Population Studies
 - Differences in development⁵⁻⁷
- Texas ecoregions^{8,9}
 - Differences in soil, vegetation, and climate



https://tp.wd.texas.gov/publications/pwdpubs/pwd_p_l_w7000_1187a/media/r1.pdf



2. Hall DG (1948) The blowflies of North America. Thomas Say Foundation
 3. Jones N, Whitworth T, Marshall SA (2019) Blow flies of North America: Keys to the subfamilies and genera of Calliphoridae, and to the species of the subfamilies Calliphorinae, Luciliinae and Chrysomyinae. Can J Arthropod Identif 1-191. <https://doi.org/10.3752/cjai.2019.39>
 4. Archangeault AD (2012) Population structure of *Lucilia mexicana* Macquart 1843 (Diptera: Calliphoridae) in Texas with a discussion of colonization and genetics. Thesis, Sam Houston State University
 5. Gallagher MB, Sandhu S, Kimsey R (2010) Variation in Developmental Time for Geographically Distinct Populations of the Common Green Bottle Fly, *Lucilia sericata* (Meigen)*. J Forensic Sci 55:438-442. <https://doi.org/10.1111/j.1556-4029.2009.01285.x>
 6. Tarone AM, Picard CJ, Spiegelman C, Foran DR (2011) Population and Temperature Effects on *Lucilia sericata* (Diptera: Calliphoridae) Body Size and Minimum Development Time. J Med Entomol 48:1062-1068. <https://doi.org/10.1603/ME11004>
 7. Owings C, Spiegelman C, Tarone A, Tomberlin J (2014) Developmental variation among *Cochliomyia macellaria* Fabricius (Diptera: Calliphoridae) populations from three ecoregions of Texas, USA. Int J Legal Med 128:709
 8. Gould FW, Hoffman GO, Rechenhain CA (1960) Vegetation of Texas.
 9. Griffith G, Bryce S, Omernik J, Anne Rogers (2007) Ecoregions of Texas

Objectives

- Objective 1: Design microsatellite multiplex for *L. mexicana*
- Objective 2: Assess the population structure of *L. mexicana* across the Texas ecoregions

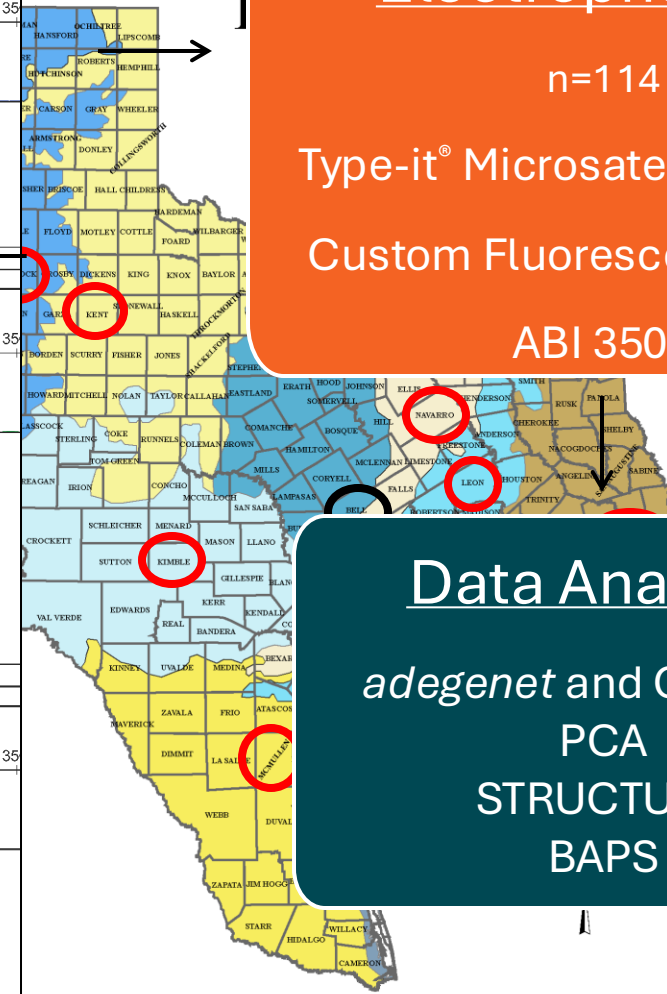
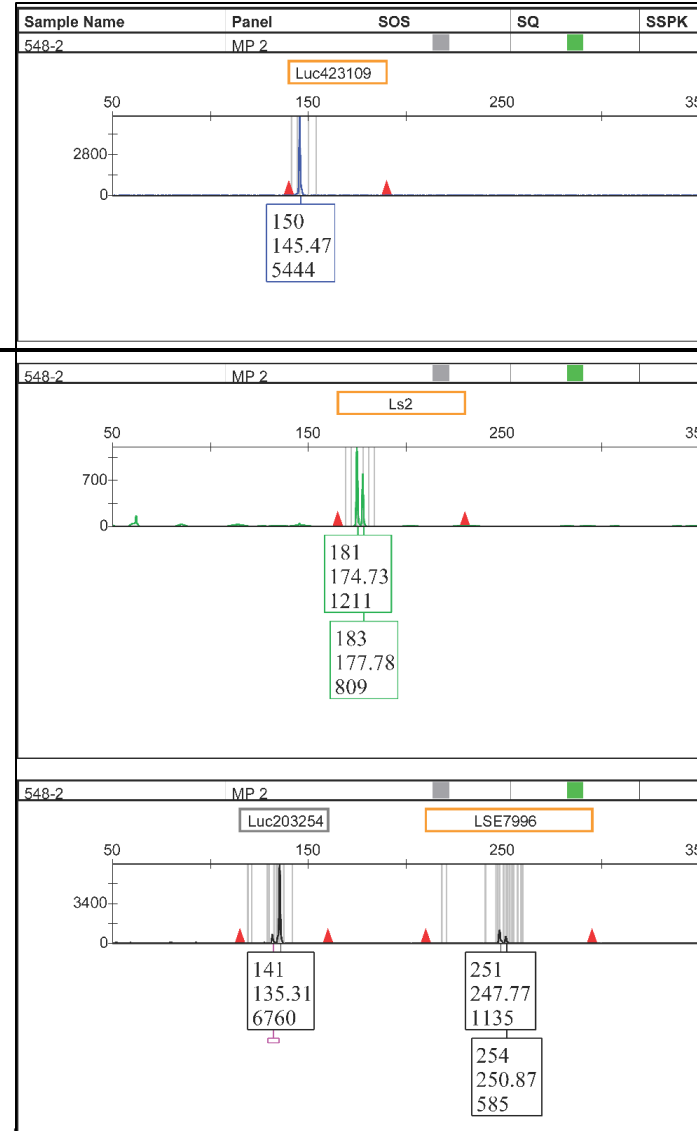
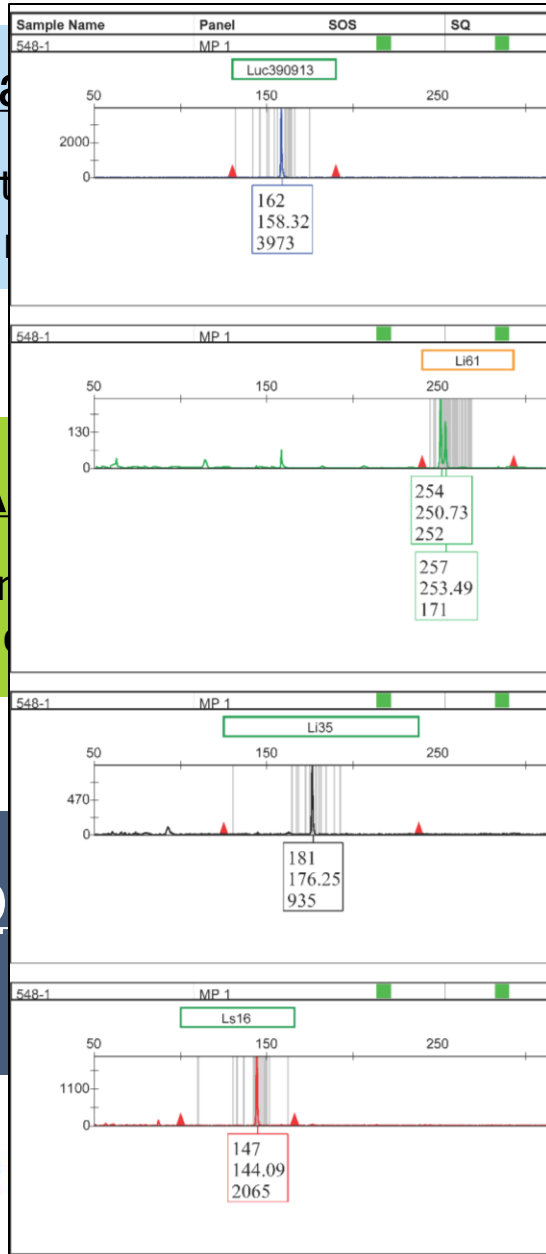


Methods

Sample
Collection
Site

DNA
Chelex and
analysis

DNA Q



PCR and Capillary Electrophoresis
n=114
Type-it® Microsatellite PCR Kit
Custom Fluorescent Primers
ABI 3500

Data Analysis
adegenet and Genepop
PCA
STRUCTURE
BAPS

Map 1

10. Florin A-B, Gyllenstrand N (2002) Isolation and characterization of polymorphic microsatellite markers in the blowflies *Lucilia illustris* and *Lucilia sericata*. *Molecular Ecology Notes* 2: 113–116. <https://doi.org/10.1046/j.1471-8286.2002.00165.x>
 11. An H-E, Do DT, Lee, et al (2021) Development of the new microsatellite markers of *Lucilia sericata* (Diptera: Calliphoridae) from Korea. *Mol Biol Rep* 48:8245–8248. <https://doi.org/10.1007/s11033-021-06750-x>
 12. Megécz E, Pech N, Gilles A, et al (2014) QDD version 3.1: a user-friendly computer program for microsatellite selection and primer design revisited: experimental validation of variables determining genotyping success rate. *Molecular Ecology Resources* 14:1302–1313. <https://doi.org/10.1111/1755-0998.12271>

Markers

Name	Forward and Reverse Primers 5'-3' Sequence	Final Reaction Concentration (μM)	Size (bp)*	Alleles	H_o/H_e	Motif^	Panel and Dye
Li35 ¹⁰	TGTCTTGTTCAATTTGATGCTTG	0.4	130-200	12	0.32/0.67	TA ₂ (CATA) ₂ TA ₂ (CA)TA ₃	1-NED
	TTCATACTTCTCTATTATTATATTTT						
Li61 ¹⁰	CCAATTACAACTTACATCC	0.1	245-275	19	0.67/0.89	-	1-VIC
	GGAATTGTTATAATTTAATT						
Ls2 ¹⁰	TCCCACGTCAACAACAAAA	0.075	170-190	6	0.45/0.70	(CAA) ₂ TAA(CCA)CAA ₃	2-VIC
	CTTGTCGAACCCTGAAGGAA						
Ls16 ⁺¹⁰	TGTAAATCATTTTAGAGAAATTCA	0.2	130-170	14	0.30/0.64	-	1-PET
	TTTGTTGTTTTCTCTGACCAA						
LSE-7996 ⁺¹¹	GTCTCTTATTGTGGCATCAT	0.075	220-270	14	0.24/0.80	-	2-NED
	GGTGAATGCCGTTATAA						
Luc203254	TTGTAAAACTCGACCACAC	0.1	120-150	8	0.21/0.53	AC ₂ TA(TGT) ₂	2-NED
	TTTGATTAGCGTGAGGAAATGA						
Luc390913	CAAGAGTTGATTGGAATACCG	0.05	130-180	14	0.52/0.59	GGAC ₃	1-6-FAM
	GGCCACTAGCTTGATGTACG						
Luc423109	AATGATTGTCAACAACCACTGG	0.05	140-160	5	0.15/0.14	(CGTC) ₃	1-6-FAM
	GAACGAAGGTTTGGCATCAGC						

*size is based on sequencing results

^ motif from Positive Control (Huntsville), (-) mononucleotide repeat (s)

+ Departure from Hardy-Weinberg

No linkage detected between the loci



Pairwise F_{st}

	Blackland Prairies	Gulf Coast Prairies and Marshes	Pineywoods	Cross Timbers North	Trans-Pecos	Cross Timbers South	Post Oak Savannah	High Plains	Rolling Plains	Edwards Plateau
Gulf Coast	0.1903									
Pineywoods	0.0005	0.3103								
Cross Timbers N	-0.0233	0.2167	-0.0038							
Trans-Pecos	0.2791	0.2845	0.3777	0.2395						
Cross Timbers S	0.2816	0.2507	0.4058	0.2298	0.0912					
Post Oak Savannah	-0.0026	0.266	0.0424	-0.0045	0.2566	0.2669				
High Plains	0.2599	0.2966	0.3616	0.2154	0.1148	0.007	0.2175			
Rolling Plains	0.2869	0.2956	0.3953	0.2387	0.0942	-0.0253	0.2464	-0.0046		
Edwards Plateau	0.3003	0.3024	0.4318	0.2593	0.1097	-0.015	0.278	0.0233	0.0041	
South Texas Plains	0.2538	0.0033	0.3749	0.268	0.2913	0.2355	0.323	0.3026	0.2922	0.297

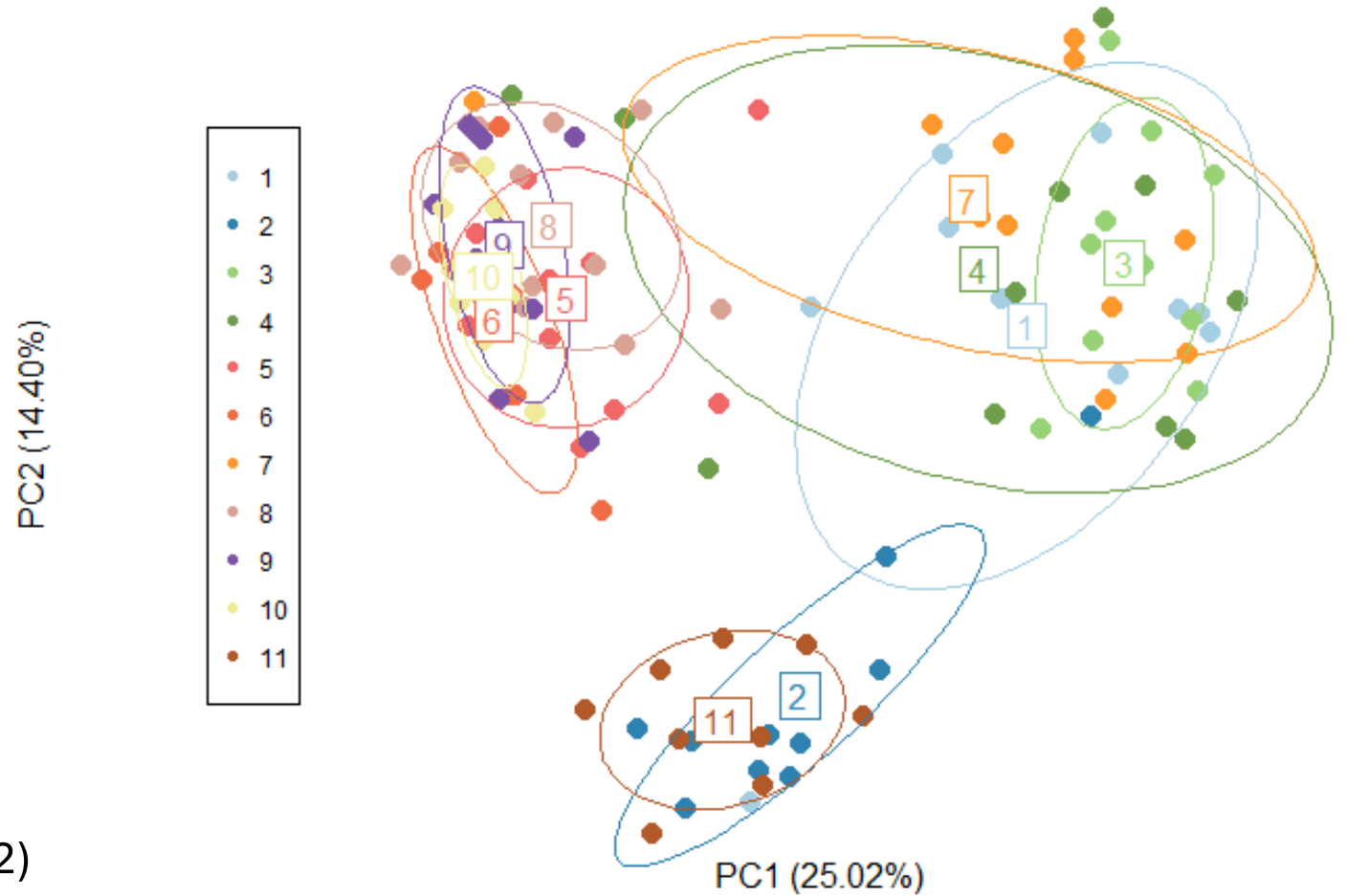


0 to 0.05: Low to no genetic differentiation
0.05 to 0.15: Moderate differentiation

0.15 to 0.25: High differentiation
Above 0.25: Very high differentiation

PCA

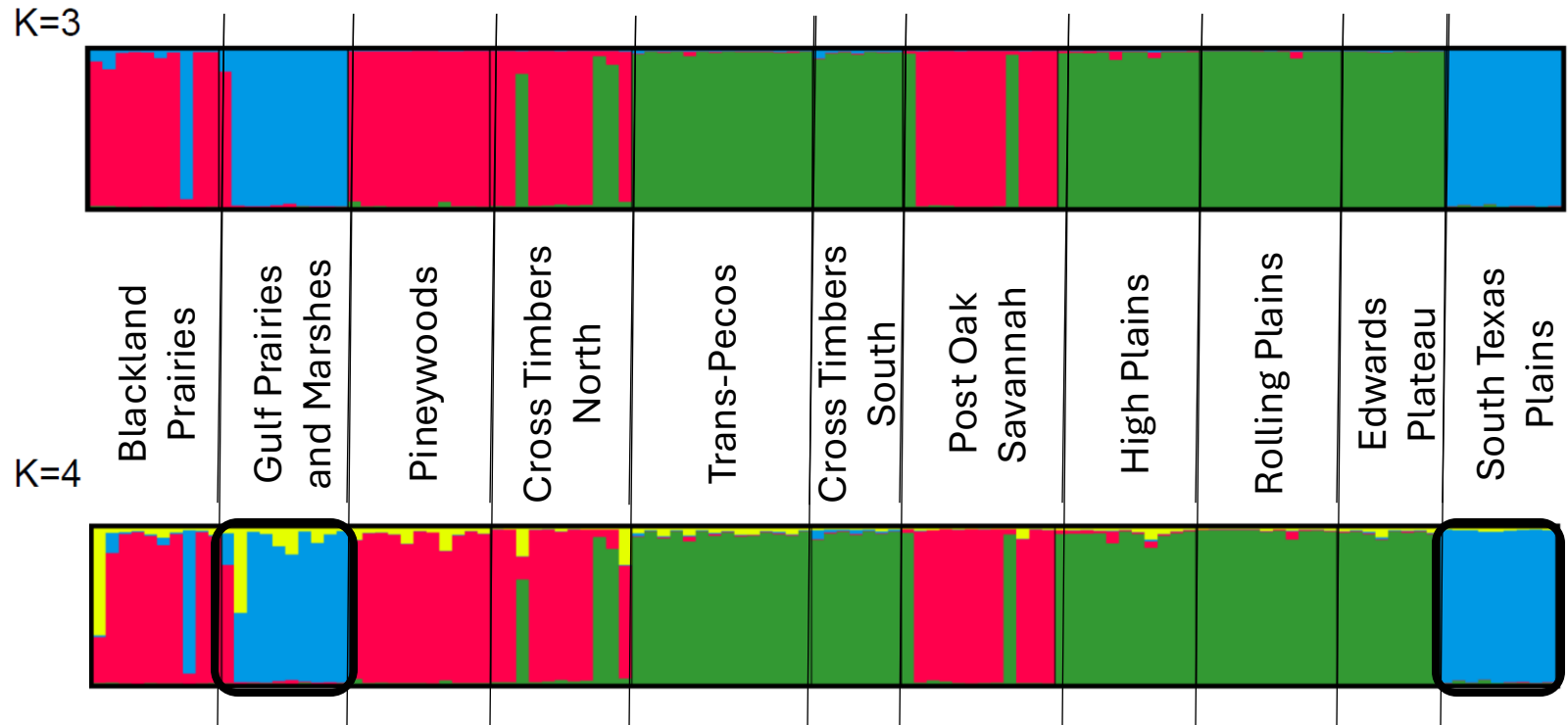
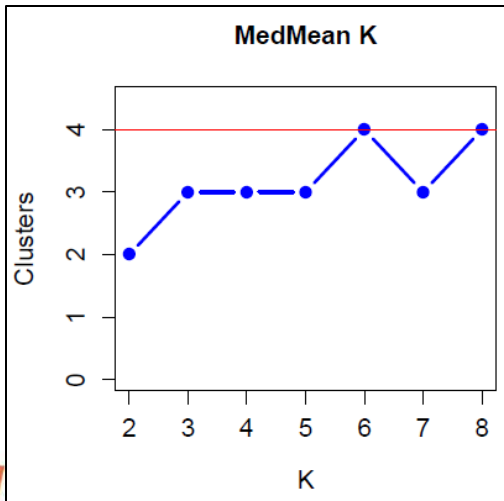
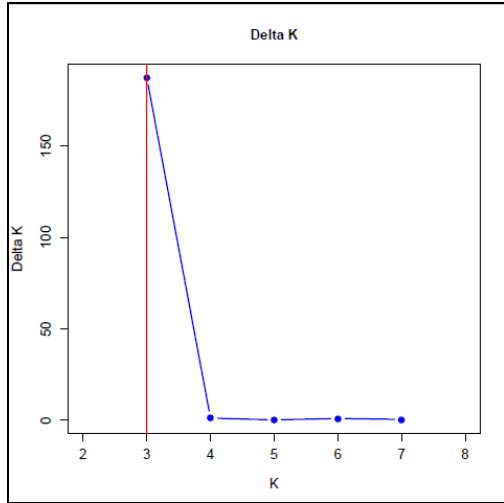
- Cluster 1
 - Blackland Prairies (1)
 - Pinewoods (3)
 - Cross Timbers North (4)
 - Post Oak Savannah (7)
- Cluster 2
 - Trans-Pecos (5)
 - Cross Timbers South (6)
 - High Plains (8)
 - Rolling Plains (9)
 - Edwards Plateau (10)
- Cluster 3
 - Gulf Coast Prairies and Marshes (2)
 - South Texas Plains (11)



STRUCTURE

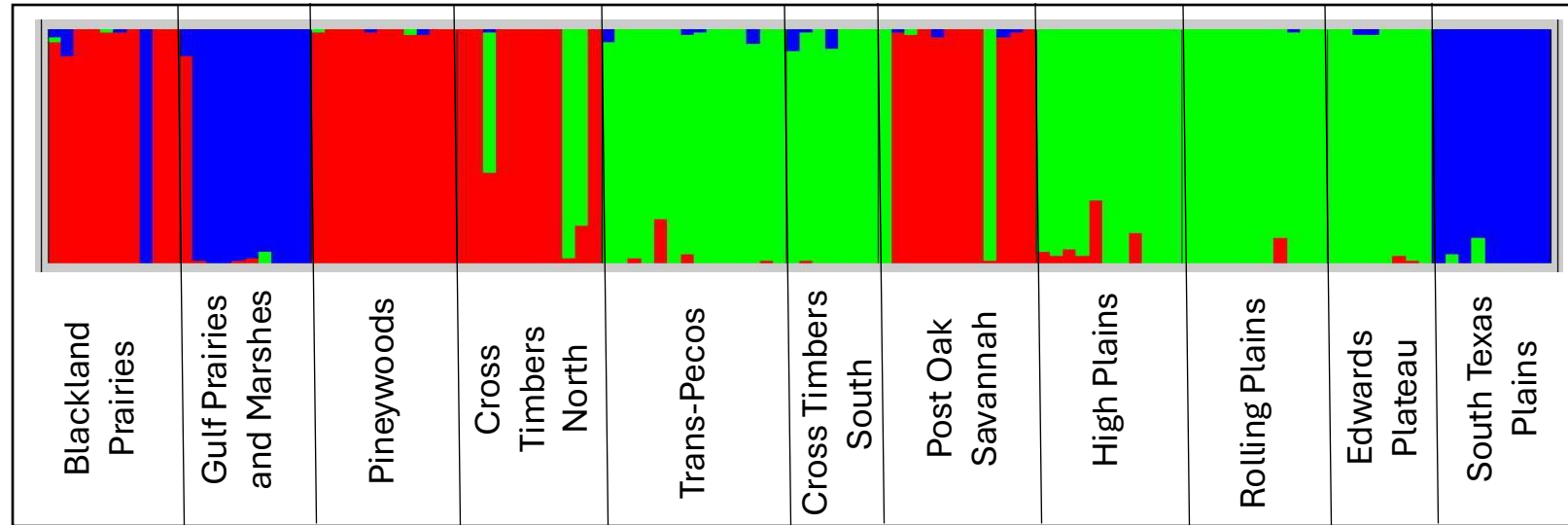
Various methods to determine best K

K=genetic clusters
3 or 4 clusters

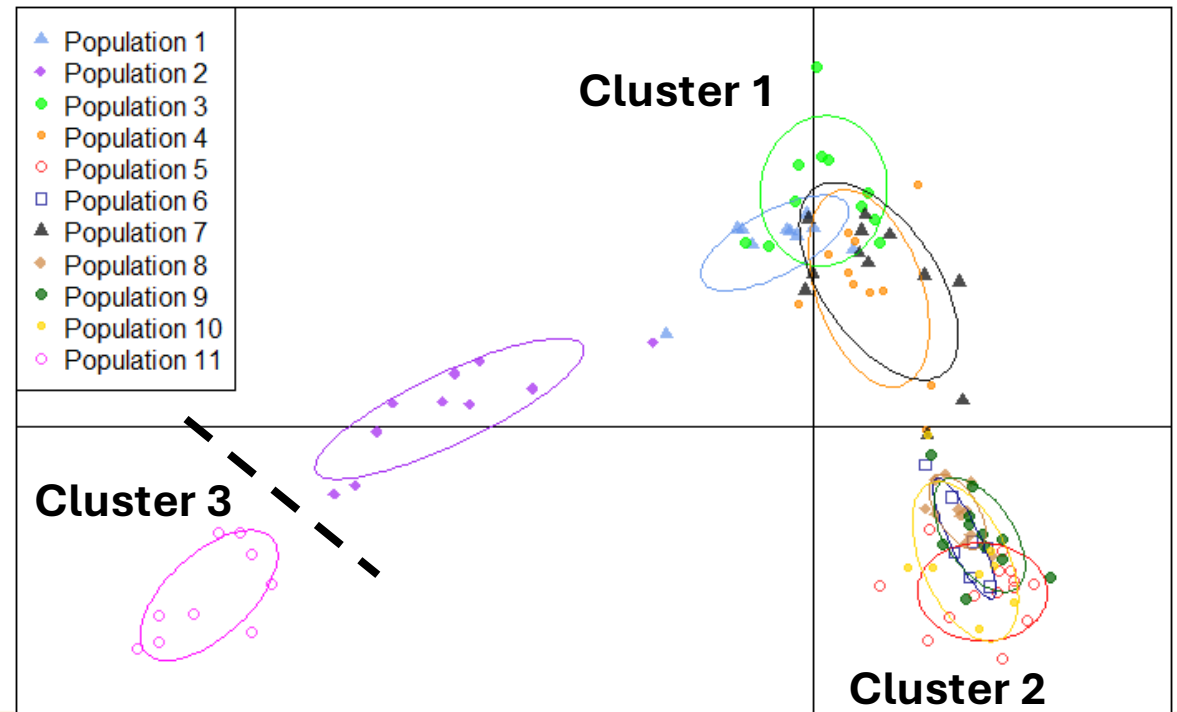


BAPS and DAPC

- BAPS
 - 3 clusters identified

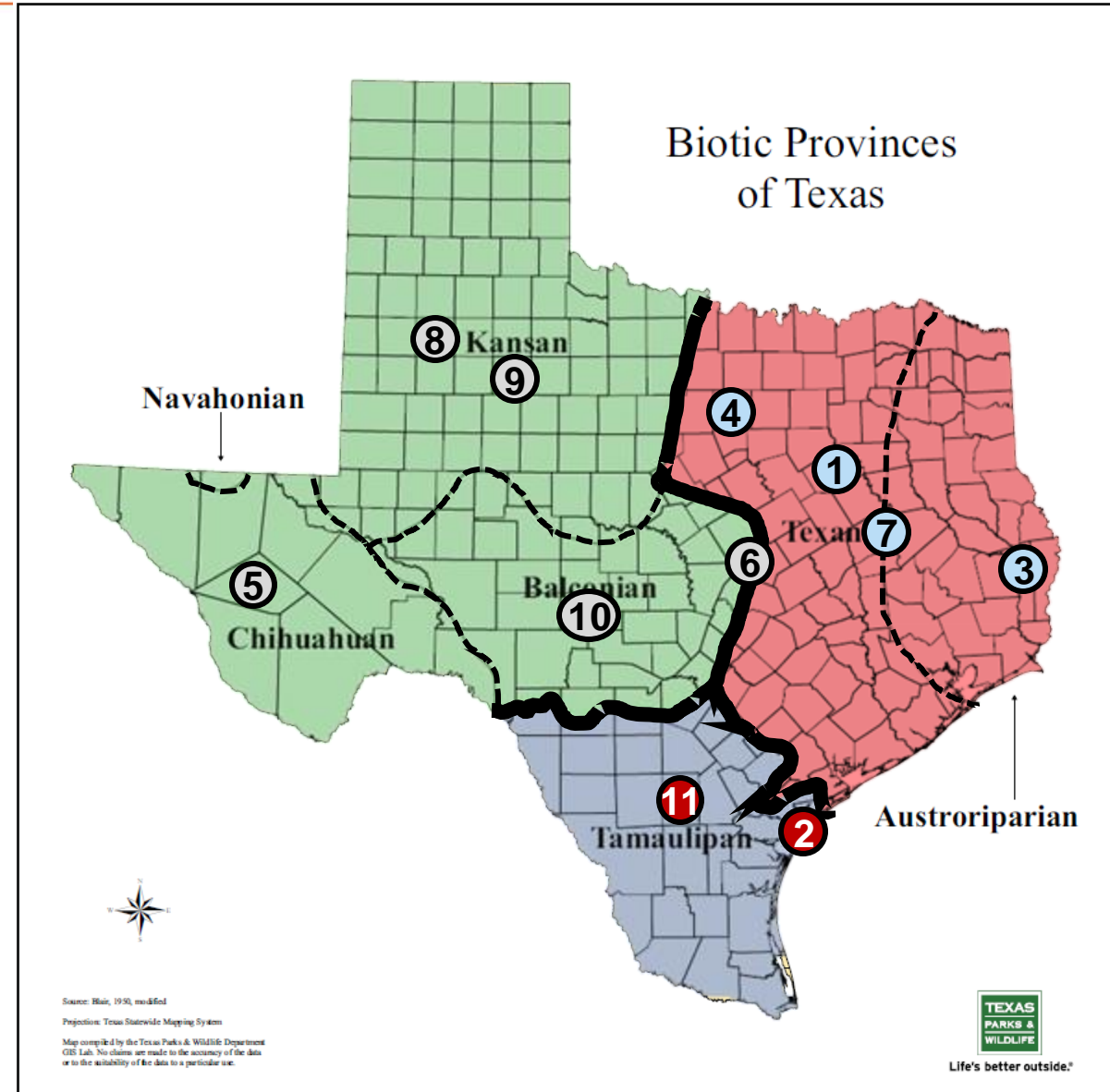


- DAPC
 - $p < 0.001$
 - Separated out Cluster 3



Correlation to Location

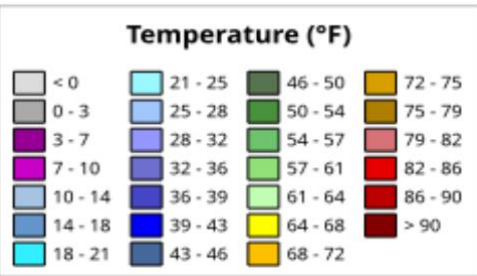
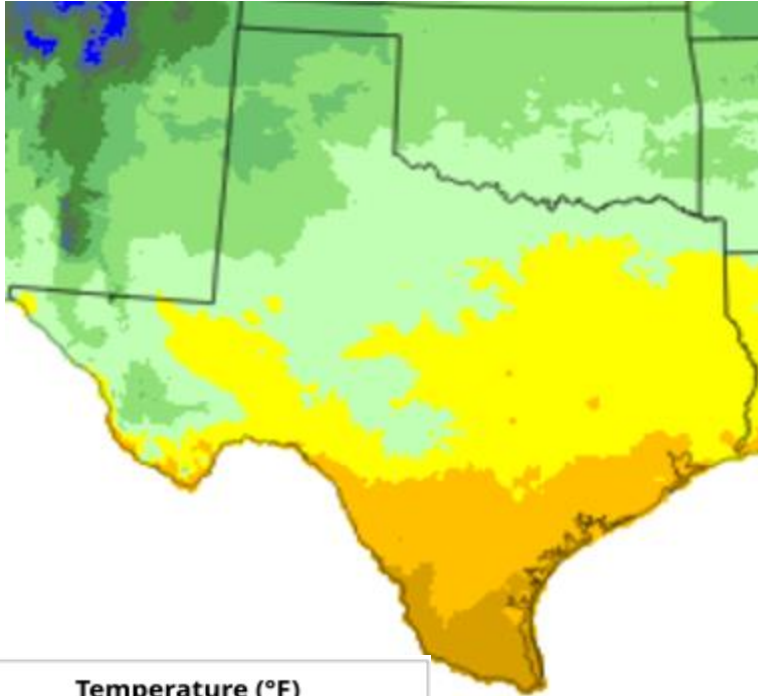
- Populations not correlated to ecoregion
- Three genetic clusters correlating with the three major biota of Texas¹³
 - Austroriparian
 - Sonoran
 - Neotropical
- Experience different temperature and precipitation



https://tpwd.texas.gov/publications/pwdpubs/media/pwd_mp_e0100_1070ae_08.pdf

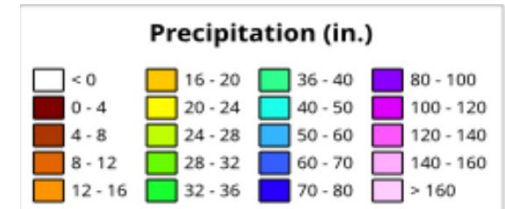
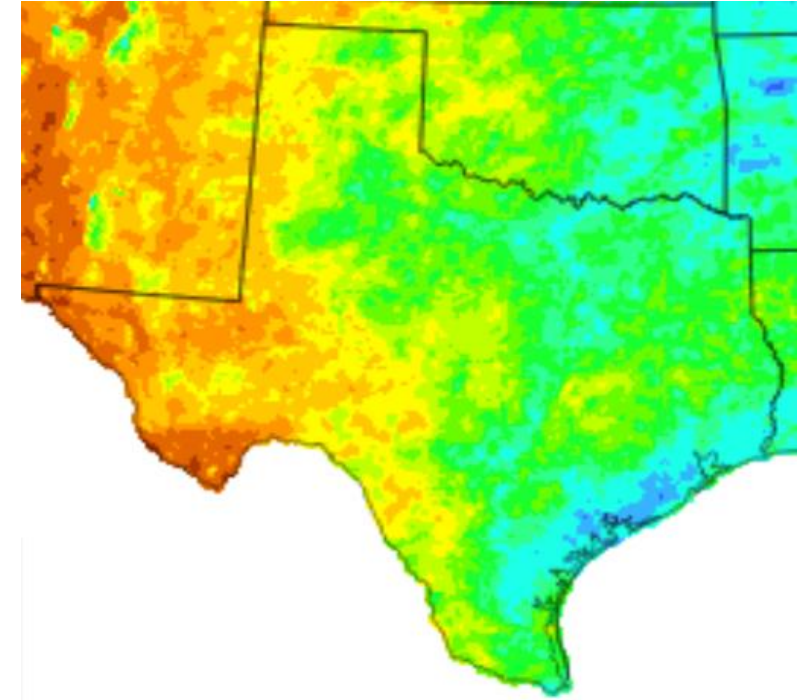
Temperature and Precipitation

Mean Temperature for 2010

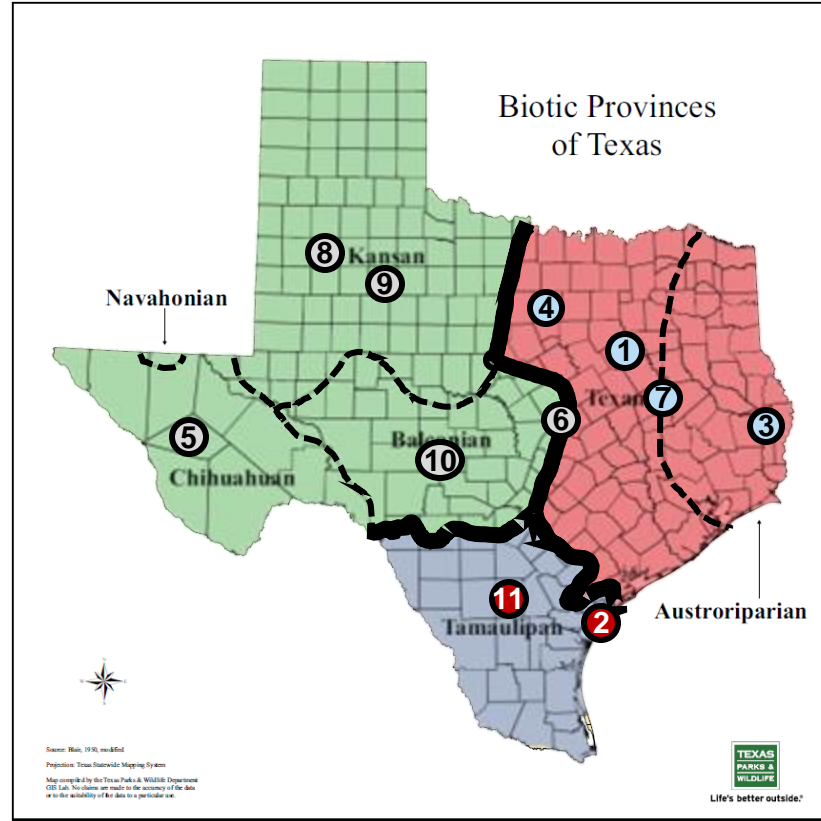


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Total Precipitation for 2010

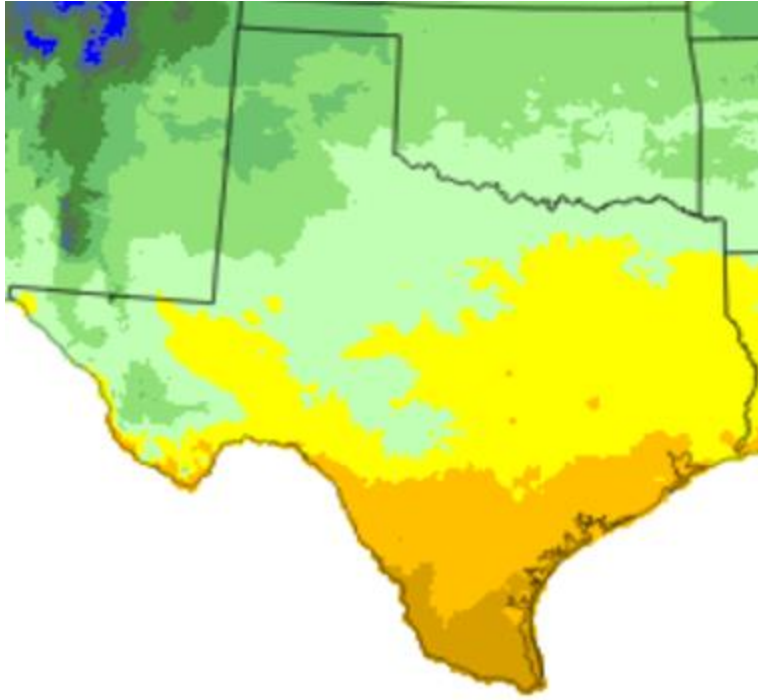


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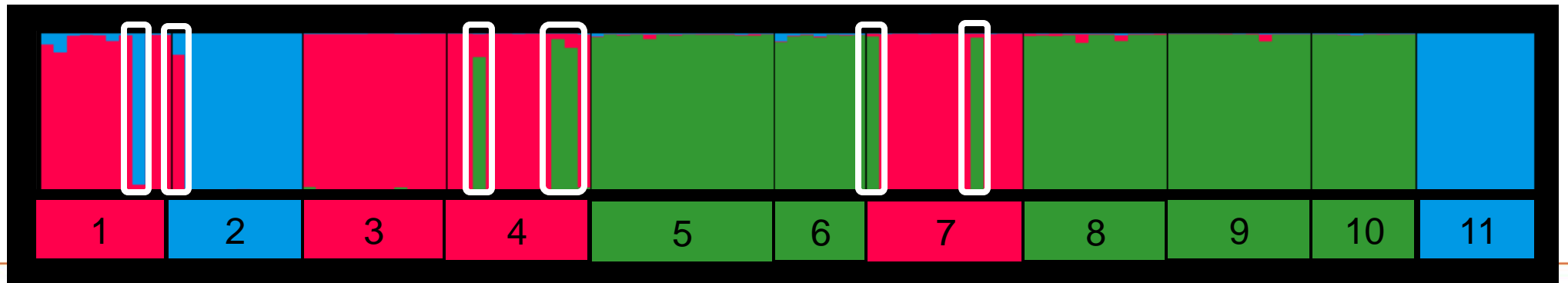
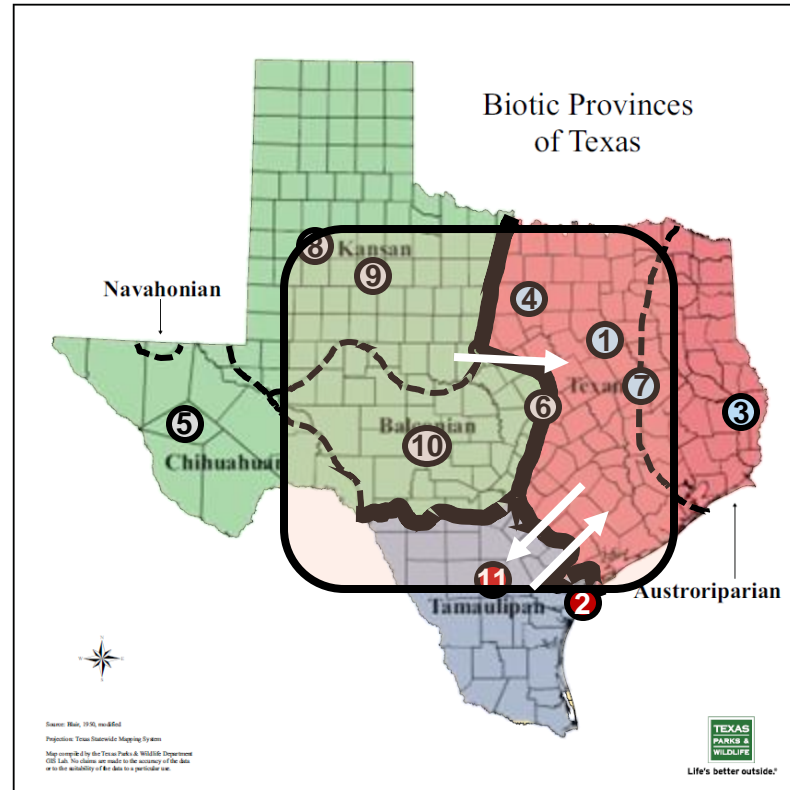
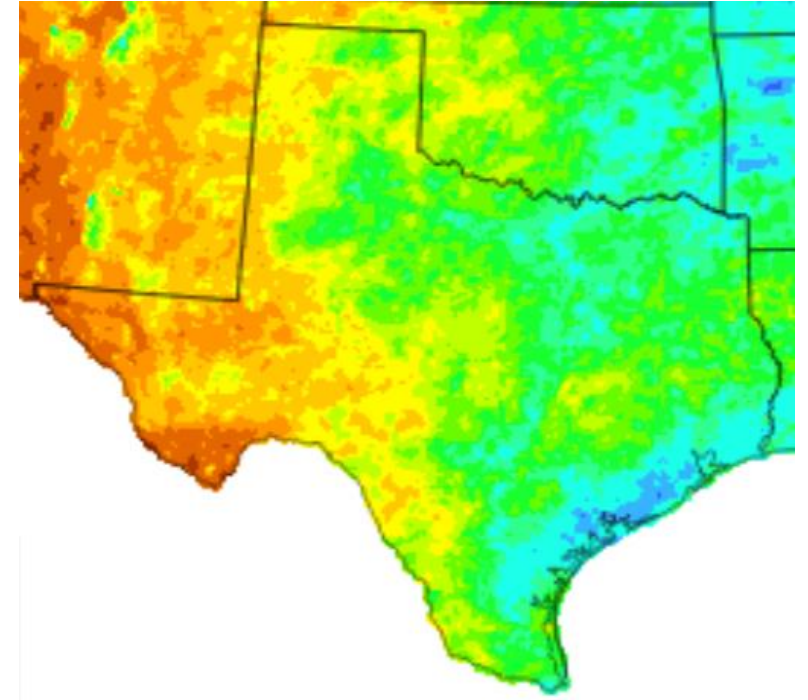


Transition Area and Migrants

Mean Temperature for 2010



Total Precipitation for 2010



June 19, 2025

July 1, 2025

July 4, 2025

July 7, 2025

DNA Barcoding

L. mexicana Population
Estimated Location
Post-mortem Interval



Conclusion

- Populations not differentiated based on ecoregions
- Clusters correlated with the three major biota of Texas
 - Temperature and precipitation gradients
- Different temperatures could lead to different development requirements and thresholds
- Future studies include the optimization of the panel
 - Additional informative markers
- Examine more recently collected *Lucilia mexicana*
 - Effects of climate change in the past 10+ years



Acknowledgements



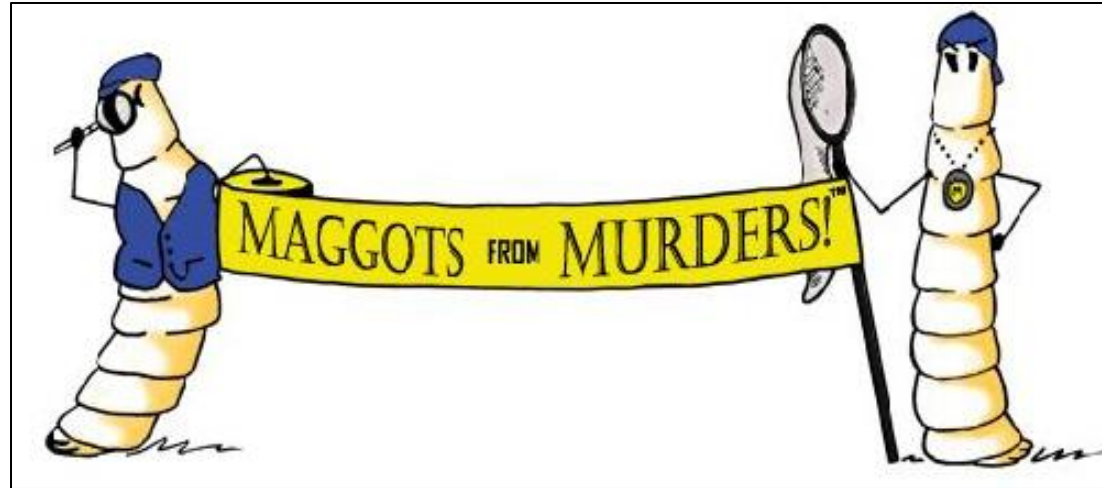
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