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# When and “Y” to Screen with QIAGEN Investigator Casework GO!

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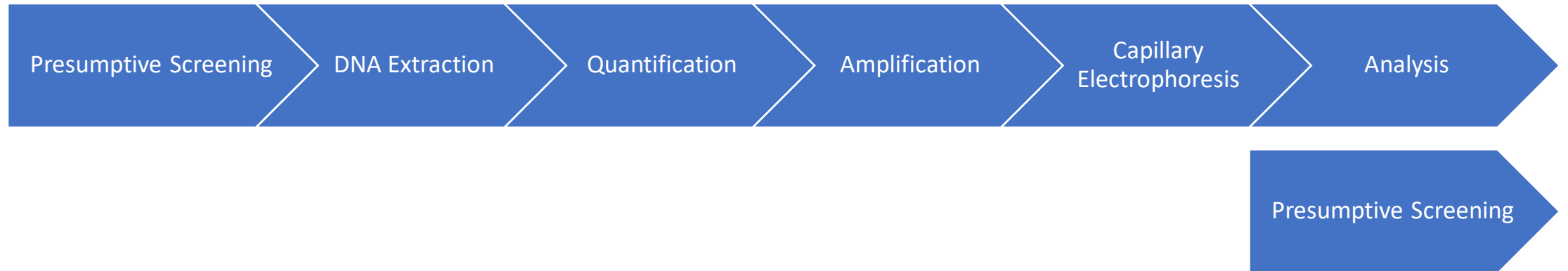
# Disclaimer

This work was supported by a collaboration with QIAGEN. The opinions, findings, and conclusions expressed in this presentation are those of the authors and do not necessarily reflect those of QIAGEN.

# Standard Sexual Assault Evidence Processing



# Y-Screening

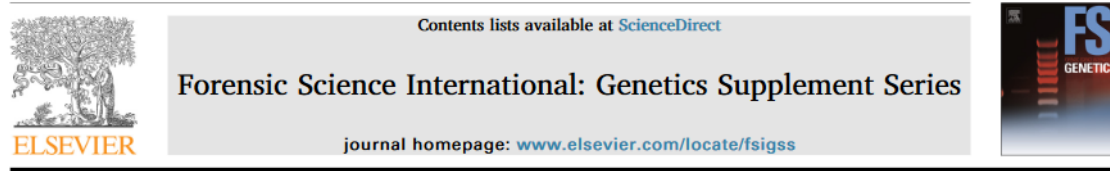


# QIAGEN Investigator Casework GO!



- **Lysis** kit designed for direct amplification workflow along with Investigator<sup>®</sup> 24plex GO! kit
- Compatible with quantification or amplification for STR analysis

# Direct PCR



How the Investigator Casework GO! Kit provides sensitive, fast and robust direct amplification of low copy number samples



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Validation Report

Developmental Validation of the Investigator<sup>®</sup>  
Casework GO! Kit

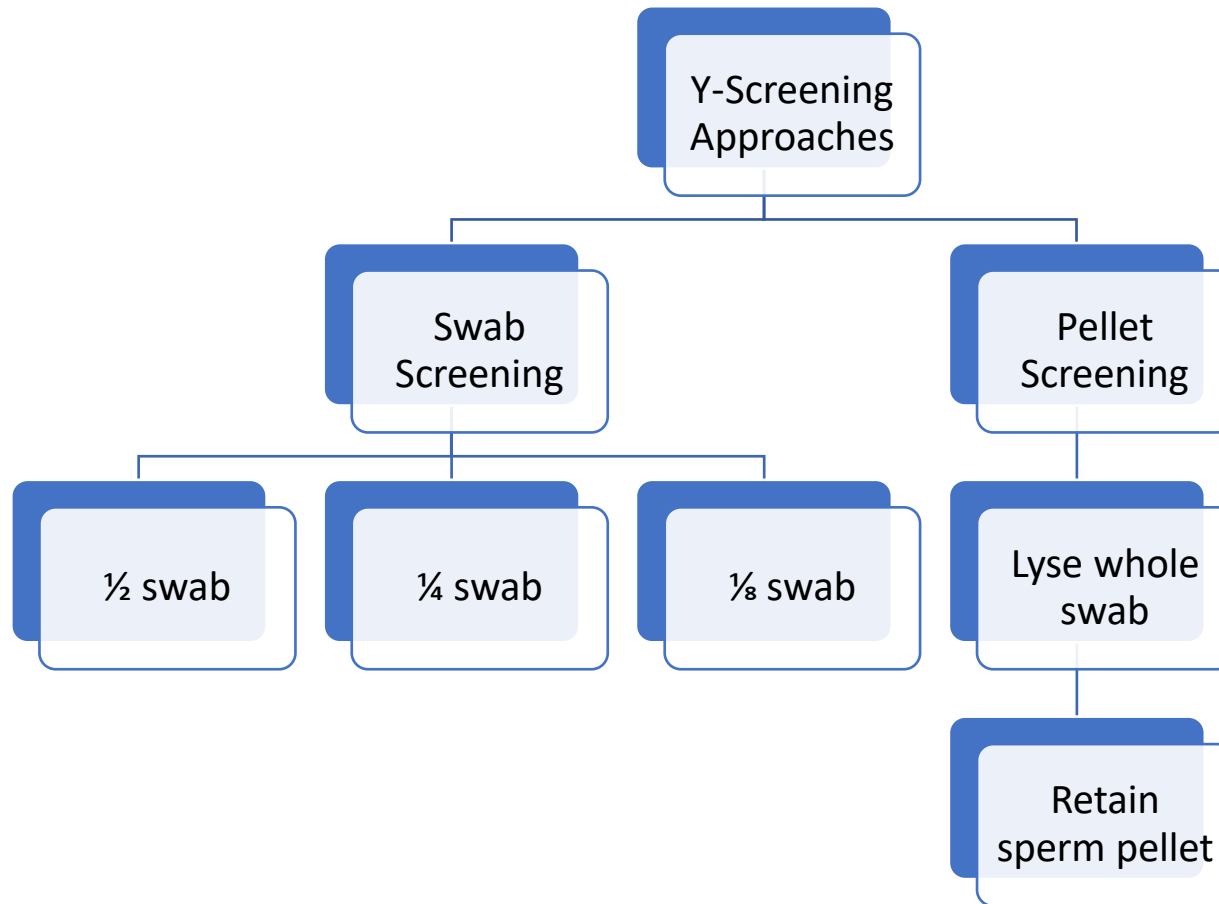
(2019):

- Blood and semen dilutions
- Direct PCR (Casework GO!) comparable to traditional method (EZ1) for blood dilutions
- 90-100% alleles recovered in semen dilutions up to 1:1000

(2022):

- Marginal DNA loss in direct PCR approach using semen dilutions
- In mock sexual assault samples, Y-STRs were recovered up to 100% after lysis with Casework GO! and differential washes
- Could not successfully perform autosomal STR analysis because of high epithelial background

# Investigator Casework GO!



# Swab Screening

Add 187  $\mu\text{L}$  Casework GO! buffer, 7  $\mu\text{L}$  ProK, 6  $\mu\text{L}$  DTT (10 mM) to sample

Vortex

Incubate 60°C for 25 min at 900 rpm

Incubate 80°C for 5 min

Transfer lysate

Quantification



# Pellet Screening

Add 290  $\mu\text{L}$  of Casework GO! buffer and 10  $\mu\text{L}$  ProK to whole swab

Incubate 60°C for 25 min at 900 rpm

Place swab into spin basket

Centrifuge for 5 min at 4500 rpm

Remove 250  $\mu\text{L}$  (epithelial fraction) from top

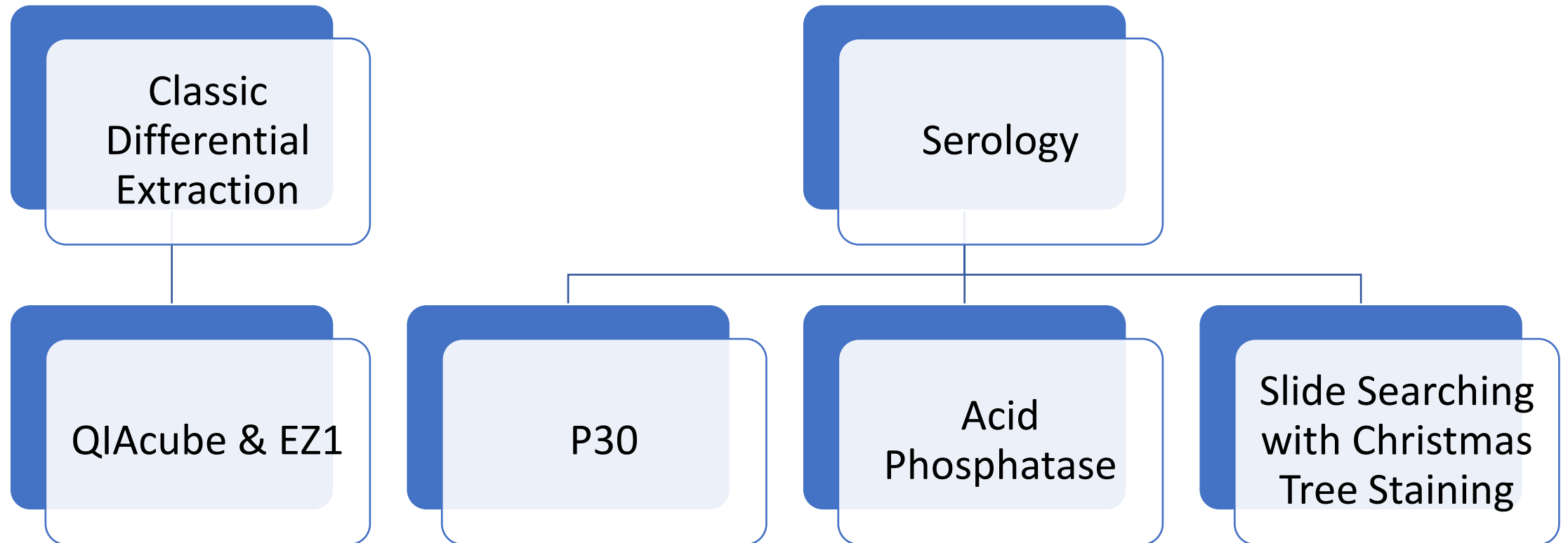
Vortex pellet and remaining 50  $\mu\text{L}$  of sample (sperm fraction)

Remove 10  $\mu\text{L}$  of pellet fraction for screening

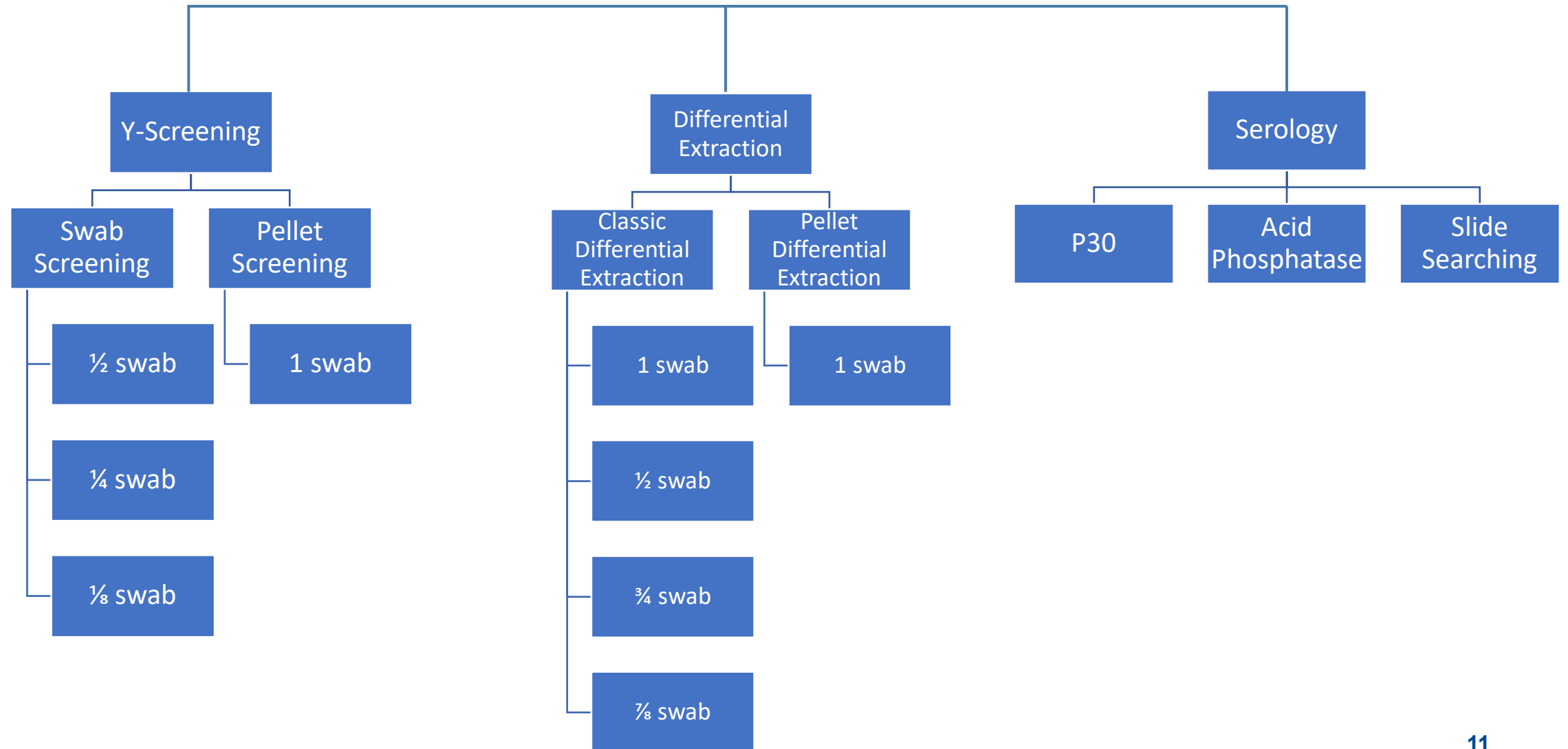
Add 1  $\mu\text{L}$  ProK and 1  $\mu\text{L}$  DTT (10 mM) to screening sample

Quantification

# Methods of Comparison



# Methods



# Samples

- Semen spiked onto vaginal swabs in triplicate at:
  - 500 ng
  - 100 ng
  - 20 ng
  - 4 ng
  - 0.8 ng
  - 0.16 ng
  - 0.032 ng
  - 0.0064 ng

# P30

Semen DNA	P30 Result
500 ng	3
100 ng	3
20 ng	2
4 ng	1
0.8 ng	0
0.16 ng	0
0.032 ng	0
0.0064 ng	0

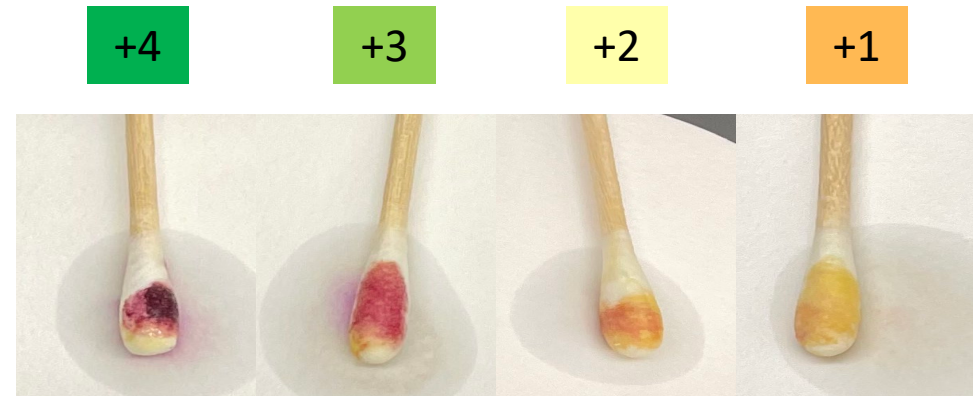
3      2      1      0



Score	Description
3	Immediate strong reaction
2	Moderate reaction
1	Weak reaction
0	No reaction; negative reaction

# Acid Phosphatase

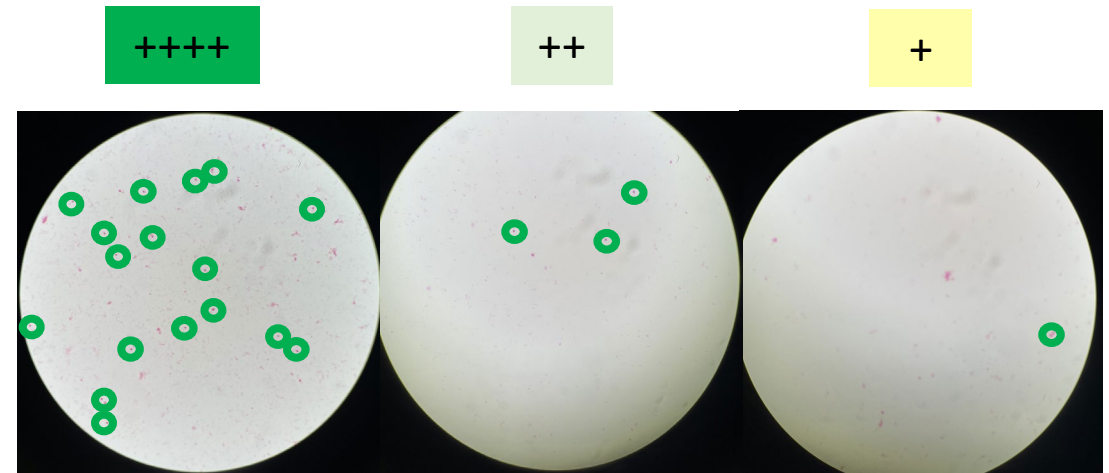
Male DNA	AP Result
500 ng	+4
100 ng	+3
20 ng	+3
4 ng	+2
0.8 ng	+2
0.16 ng	+2
0.032 ng	+1
0.0064 ng	+1



Score	Description
+4	Intense purple color develops and bleeds into test paper
+3	Intense purple color develops and develops quickly
+2	2: Slight purple color develops on and may develop slowly
+1	1: Slow pink color develops on stain material

# Slide Searching

Male DNA	Slide Searching Result
500 ng	+++
100 ng	++
20 ng	++
4 ng	+
0.8 ng	0
0.16 ng	0
0.032 ng	0
0.0064 ng	0



Score	Description
++++	Many spermatozoa in every field
+++	Many or some spermatozoa in most fields
++	Some spermatozoa in some fields, easy to find
+	Hard to find
0	No spermatozoa observed

# Male Target Quantification (ng/ $\mu$ L) & Male:Human Ratio

	Casework GO! Screening				Differential Extraction					Serology		
	½ Swab Screen	¼ Swab Screen	⅛ Swab Screen	Y Pellet Screen	Whole Swab DE	½ DE	¾ DE	⅞ DE	Y Pellet DE	P30	AP	Slide Searching
500 ng	0.28 1:30	0.53 1:9	0.32 1:9	14.89 1:23	5.80 1:1	2.42 1:1	1.50 1:1	0.82 1:1	3.12 1:1	3	4	+++
100 ng	0.04 1:1E2	0.07 1:36	0.09 1:38	3.42 1:51	2.81 1:1	0.33 1:1	0.22 1:3	0.22 1:2	0.24 1:2	3	3	++
20 ng	0.02 1:2E2	0.03 1:1E2	0.01 1:2E2	0.74 1:3E2	0.43 1:4	0.03 1:6	0.06 1:8	0.11 1:2	0.12 1:24	2	3	++
4 ng	0.003 1:2E3	0.01 1:2E2	0.01 1:4E2	0.08 1:7E2	0.04 1:17	0.01 1:21	0.004 1:27	0.02 1:32	0.02 1:21	1	2	+
0.8 ng	0.0003 1:2E4	0.002 1:4E3	0.002 1:2E3	0.04 1:4E3	0.02 1:81	0.001 1:53	0.003 1:41	0.007 1:2E2	0.001 1:2E2	0	2	0
0.16 ng	0.0001 1:4E4	0.0007 1:3E3	0.007 1:2E3	0.009 1:2E4	0.00 1:3E2	0.0004 1:23	0.0007 1:2E2	0.0005 1:5E2	0.001 1:2E2	0	2	0
0.032 ng	0.00 N/A	0.0008 1:2E3	0.001 1:5E3	0.0006 1:3E4	0.00 1:2E3	0.0002 1:2E2	0.0003 1:3E2	0.0006 1:2E2	0.0004 1:4E2	0	1	0
0.0064 ng	0.0001 1:3E4	0.0009 1:4E3	0.0005 1:2E3	0.00007 1:2E5	0.00 N/A	0.00 N/A	0.00007 1:4E2	0.0001 1:2E2	0.0003 N/A	0	1	0



# Results: STR Mixture Analysis

Range of log( <i>Likelihood Ratios</i> ) from Spiked Swabs					
	500 ng	100 ng	20 ng	4 ng	0.8 ng
Classic Differential	19-24	24	21-37	5-11	0-1
Y Pellet Differential	24	24	2-14	4-9	0-3

# Post-Coital Swab Testing

## Post-coital swabs collected at:

- 6 hr
- 12 hr
- 24 hr
- 48 hr

## Screening

- Serology
- Swab Screening with Casework GO!
- Pellet Screening with Casework GO!

## Extraction

- Classic Differential Extraction
- Y Pellet Differential Extraction

## Mixture Analysis with STRmix™ 2.9.1

# Male Target Quantification (ng/ $\mu$ L) & Male:Human Ratio

	Casework GO! Screening			Differential Extraction		Serology	
	¼ Swab Screen	⅛ Swab Screen	Y Pellet Screen	Whole Swab DE	Y Pellet DE	AP	Slide Searching
6 hr	0.14 1:82	0.22 1:1E2	0.81 1:1E3	5.16 1:1	3.32 1:1	3	+++
12 hr	0.03 1:1E3	0.07 1:2E2	0.21 1:4E2	0.86 1:1	0.42 1:2	3	+
24 hr	0.01 1:1E3	0.02 1:9E2	0.05 1:3E4	0.14 1:3	4.48 1:24	3	+
48 hr	0.02 1:3E3	0.02 1:5E3	0.16 1:5E4	0.80 1:5	0.33 1:21	2	0

# Results: Post-Coital STR Mixture Analysis

Range of $\log(\text{Likelihood Ratios})$ from Post-Coital Swabs				
	6 hr	12 hr	24 hr	48 hr
Classic Differential	25	25	12-25	8-25
Y Pellet Differential	25	25	24-25	17-25

# Conclusions

- Y Pellet screening with QIAGEN Casework GO! was the most successful method.
  - Screening most closely matched results observed after full differential extraction.
- Higher Human:Male ratios in screened samples were indicative of epithelial crossover from lack of differential washes.
  - Can still successfully screen for presence of male DNA.
- Results with genuine post-coital showed successful Y Pellet differential workflow.
  - LRs within the same orders of magnitude between Y Pellet and Classic differential extraction.

# Acknowledgements



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# Questions?

Julia Wang, MS

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